

## PATENT COOPERATION TREATY

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## NOTIFICATION OF ELECTION

(PCT Rule 61.2)

From the INTERNATIONAL BUREAU

To:

Commissioner  
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United States Patent and Trademark  
Office, PCT  
2011 South Clark Place Room  
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ETATS-UNIS D'AMERIQUE  
in its capacity as elected Office

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International filing date (day/month/year) 11 August 2000 (11.08.00)	Priority date (day/month/year) 12 August 1999 (12.08.99)
Applicant DRIEU, Katy et al	

1. The designated Office is hereby notified of its election made:

☒ in the demand filed with the International Preliminary Examining Authority on:  
12 March 2001 (12.03.01)

☐ in a notice effecting later election filed with the International Bureau on:  
\_\_\_\_\_

2. The election ☒ was  
☐ was not

made before the expiration of 19 months from the priority date or, where Rule 32 applies, within the time limit under Rule 32.2(b).

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The International Bureau of WIPO 34, chemin des Colombettes 1211 Geneva 20, Switzerland Facsimile No.: (41-22) 740.14.35	Authorized officer Maria Kirchner Telephone No.: (41-22) 338.83.38
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## PATENT COOPERATION TREATY

From the INTERNATIONAL BUREAU

**PCT**

**NOTICE INFORMING THE APPLICANT OF THE  
COMMUNICATION OF THE INTERNATIONAL  
APPLICATION TO THE DESIGNATED OFFICES**

(PCT Rule 47.1(c), first sentence)

**To:**

**TSAO, Y., Rocky  
Fish & Richardson P.C.  
225 Franklin Street  
Boston, MA 02110-2804  
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Date of mailing (day/month/year) 22 February 2001 (22.02.01)		FISH & RICHARDS BOSTON OFFICE	
Applicant's or agent's file reference 000215 WO1		IMPORTANT NOTICE	
International application No. PCT/US00/22174	International filing date (day/month/year) 11 August 2000 (11.08.00)	Priority date (day/month/year) 12 August 1999 (12.08.99)	
Applicant SOCIETE DE CONSEILS DE RECHERCHES ET D'APPLICATIONS SCIENTIFIQUES, S.A.S. et al			

1. Notice is hereby given that the International Bureau has communicated, as provided in Article 20, the international application to the following designated Offices on the date indicated above as the date of mailing of this Notice:

AU, KP, KR, US

In accordance with Rule 47.1(c), third sentence, those Offices will accept the present Notice as conclusive evidence that the communication of the international application has duly taken place on the date of mailing indicated above and no copy of the international application is required to be furnished by the applicant to the designated Office(s).

- 2. The following designated Offices have waived the requirement for such a communication at this time:**

AE,AG,AL,AM,AP,AT,AZ,BA,BB,BG,BR,BY,BZ,CA,CH,CN,CR,CU,CZ,DE,DK,DM,DZ,EA,EE,EP,ES,  
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no communication will be made to those Offices only upon their request. Furthermore, those Offices do not require the  
applicant to furnish a copy of the international application (Rule 49.1(a-bis)).

3. Enclosed with this Notice is a copy of the international application as published by the International Bureau of the WIPO. **No Declaration Required**

22 February 2001 (22.02.01) under No. WO 01/12208

**REMINDER REGARDING CHAPTER II (Article 31(2)(a) and Rule 54.2)**

If the applicant wishes to postpone entry into the national phase until 30 months (or later in some Offices) from the priority date, a demand for international preliminary examination must be filed with the competent International Preliminary Examining Authority before the expiration of 19 months from the priority date.

**It is the applicant's sole responsibility to monitor the 19-month time limit.**

**Note that only an applicant who is a national or resident of a PCT Contracting State which is bound by Chapter II has the right to file a demand for international preliminary examination.**

### REMINDER REGARDING ENTRY INTO THE NATIONAL PHASE (Article 22 or 39(1))

If the applicant wishes to proceed with the international application in the **national phase**, he must, within 20 months or 30 months, or later in some Offices, perform the acts referred to therein before each designated or elected Office.

For further important information on the time limits and acts to be performed for entering the national phase, see the Annex to Form PCT/IB/301 (Notification of Receipt of Record Copy) and Volume II of the PCT Applicant's Guide.

<p><b>The International Bureau of WIPO</b>  <b>34, chemin des Colombettes</b>  <b>1211 Geneva 20, Switzerland</b></p> <p>Facsimile No. (41-22) 740.14.35</p>	<p>Authorized officer</p> <p><b>J. Zahra</b></p> <p>Telephone No. (41-22) 338.83.38</p>
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**WO 01/12208 A1**

(54) Title: USE OF GINKGO EXTRACT

(57) Abstract: The present invention is directed to the use of the extract of Ginkgo biloba leaves or isolated Ginkgolide B (GKB), a component of the extract of Ginkgo biloba leaves in a method for decreasing the expression of peripheral-type benzodiazepine receptor (PBR) in cells of a patient in need thereof. Further, the present invention is directed to the use of the extract of Ginkgo biloba leaves or isolated GKB in a method for decreasing the proliferation of cancer cells in a patient. More particularly, the present invention is directed to the use of the extract of Ginkgo biloba leaves or isolated GKB in a method of decreasing cancer cell proliferation in a patient wherein the cancer cell is human breast cancer cell. Even more particularly, the present invention is directed to the use of the extract of Ginkgo biloba leaves or isolated GKB in method of decreasing cancer cell proliferation in a patient wherein the cancer cell is of the aggressive and invasive phenotype and expresses high levels of PBR in comparison to non-aggressive cancer cell.

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## USE OF GINKGO EXTRACT

Background of the Invention

The present invention is directed to the use of the extract of *Ginkgo biloba* leaves or  
5 isolated ginkgolide B (GKB), a component of the extract of *Ginkgo biloba* leaves in a method  
for decreasing the expression of peripheral-type benzodiazepine receptor (PBR) in cells of a  
patient in need thereof. Further, the present invention is directed to the use of the extract of  
*Ginkgo biloba* leaves or isolated GKB in a method for decreasing the proliferation of cancer  
cells in a patient in need thereof. More particularly, the present invention is directed to the  
10 use of the extract of *Ginkgo biloba* leaves or isolated GKB in a method of decreasing cancer  
cell proliferation in a patient in need thereof wherein said cancer cell is human breast cancer  
cells. Even more particularly, the present invention is directed to the use of the extract of  
*Ginkgo biloba* leaves or isolated GKB in a method of decreasing cancer cell proliferation in a  
patient in need thereof wherein said cancer cell are of the aggressive and invasive  
15 phenotype and expresses high levels of PBR in comparison to non-aggressive cancer cells.

In another aspect, the present invention is directed to the use of extract of *Ginkgo biloba* leaves to decrease the expression of thirty-five (35) gene products as is further detailed, hereinbelow.

It is preferred that a particular formulation of *Ginkgo biloba* leaves extract known as  
20 EGB 761® (a product of IPSEN, Paris, France) be a constituent in a composition or used in  
a method of the present invention.

*Ginkgo biloba* is one of the most ancient trees and extracts from its leaves have been used in traditional medicine for several hundred years. There are numerous studies describing the beneficial effects of *Ginkgo biloba* extracts on patients with disturbances in  
25 vigilance, memory, and cognitive functions associated with aging and senility, and on those  
with all types of dementias, mood changes, and the ability to cope with daily stressors. A  
standardized extract of *Ginkgo biloba* leaves, termed EGB 761®, has been used in most of  
these studies. This extract is also known to have cardioprotective effects (DeFeudis F.V.  
*Ginkgo biloba* extract (EGB 761®): from chemistry to clinic. Ullstein Medical, Wiesbaden,  
30 Germany. 400 pp. 1998; Tosaki, A., Droy-Lefaix, M.T., Pali, T., and Das, D.K., Free Rad.  
Biol. Med., 14: 361-370, 1993). These effects have been attributed, at least in part, to the  
free radical scavenging properties of EGB761®, probably due to the presence of flavonoid  
or terpenoid constituents in the extract. Recent *in vivo* and *in vitro* studies demonstrated that

the terpene constituents of EGB 761®, ginkgolides and bilobalide, have anti-oxidant properties (Pietri, S., Maurelli, E., Drieu, K., and Culcasi, M., *J. Mol. Cell. Cardiol.*, 29: 733-742, 1997; Yao, Z., Boujrad, N., Drieu, K., and Papadopoulos, V., *Adv. Ginkgo Biloba Res.* 7: 129-138, 1998). Other studies of EGB 761® have reported medicinal value of the product  
5 in the treatment of a variety of clinical disorders including cerebrovascular and peripheral vascular insufficiencies associated with aging and senility. See e.g., *Ginkgo biloba Extract (EGB 761®) Pharmacological Activities and Clinical Applications*, DeFeudis, F.V., Eds, Elsevier, 1991; and Ullstein Medical 1998, *Ginkgo biloba extract (EGB 761®)*, Eds. Wiesbaden, DeFeudis, F.V. The extract contains 24% ginkgo-flavone glycosides, 6%  
10 terpene lactones (ginkgolides and bilobalide), about 7% proanthocyanidins and several other constituents. See Boralle, N., et al., In: *Ginkgolides, Chemistry, Biology, Pharmacology and Clinical perspectives*, Ed: Braquet, P., J.R. Prous Science Publishers, 1988.

Tumor progression is a multi-step process in which normal cells gradually acquire  
15 more malignant phenotypes, including the ability to invade tissues and form metastases, the primary cause of mortality in breast cancer. During this process, the "aberrant" expression of a number of gene products may be the cause or the result of tumorigenesis. Considering that the first step of tumor progression is cell proliferation, it can be proposed that tumorigenesis and malignancy are related to the proliferative potential of tumoral cells.

20 Studies in a number of tumors such as rat brain containing glioma tumors (Richfield, E.K. et al. (1988) *Neurology* 38:1255-1262), colonic adenocarcinoma and ovarian carcinoma (Katz, Y. et al. (1988) *Eur. J. Pharmacol.* 148:483-484 and Katz, Y. et al. (1990) *Clinical Sci.* 78:155-158) have shown an abundance of peripheral-type benzodiazepine receptors (PBR) compared to normal tissue. Moreover, a 12-fold increase in PBR density relative to  
25 normal parenchyma, was found in human brain glioma or astrocytoma (Cornu, P. et al. (1992) *Acta Neurochir.* 199:146-152). The authors suggested that PBR densities may reflect the proliferative activity of the receptor in these tissues. Recently, the involvement of PBR in cell proliferation was further shown (Neary, J.T. et al. (1995) *Brain Research* 675:27-30; Miettinen, H. et al. (1995) *Cancer Research* 55:2691-2695), and its expression of  
30 human astrocytic tumors was found to be associated with tumor malignancy and proliferative index (Miettinen, H. et al. *supra*; Alho, H. (1994) *Cell Growth Different.* 5:1005-1014). Further studies have shown that PBR receptors are abundant in human

glioblastomas (Broaddus, W.C., et al., *Brain Research*, Vol. 518:199-208, 1990; and Pappata, S., et al., *J. Nuclear Med.*, 32:1608-1610, 1991).

PBR is an 18-kDa protein discovered as a class of binding sites for benzodiazepines distinct from the GABA neurotransmitter receptor (Papadopoulos, V. (1993) *Endocr. Rev.* 5 14:222-240). PBR are extremely abundant in steroidogenic cells and found primarily on outer mitochondrial membranes (Anholt, R. et al. (1986) *J. Biol. Chem.* 261:576-583). PBR is thought to be part of the multimeric complex composed of the 18-kDa isoquinoline-binding protein and the 34-kDa pore-forming voltage-dependent anion channel protein, preferentially located on the outer/inner mitochondrial membrane contact sites (McEnery, M. W. et al. 10 *Proc. Natl. Acad. Sci. U.S.A.* 89:3170-3174; Garnier, M. et al. (1994) *Mol. Pharmacol.* 45:201-211; Papadopoulos, V. et al. (1994) *Mol. Cell. Endocr.* 104:R5-R9). Drug ligands of PBR, upon binding to the receptor, stimulate steroid synthesis in steroidogenic cells *in vitro* (Papadopoulos, V. et al. (1990) *J. Biol. Chem.* 265:3772-3779; Ritta, M.N. et al. (1989) *Neuroendocrinology* 49:262-266; Barnea, E.R. et al. (1989) *Mol. Cell Endocr.* 64:155-159; 15 Amsterdam, A. and Suh, B. S. (1991) *Endocrinology* 128:503-510; Yanagibashi, K. et al. (1989) *J. Biochem. (Tokyo)* 106:1026-1029). Likewise, *in vivo* studies showed that high affinity PBR ligands increase steroid plasma levels in hypophysectomized rats (Papadopoulos V. et al (1997) *Steroids* 62:21-28 ). Further *in vitro* studies on isolated mitochondria provided evidence that PBR ligands, drug ligands, or the endogenous PBR 20 ligand, the polypeptide diazepam binding inhibitor (BDI) (Papadopoulos, V. et al. (1997) *Steroids* 62:21-28), stimulate pregnenolone formation by increasing the rate of cholesterol transfer from the outer to the inner mitochondrial membrane (Krueger, K. E. and Papadopoulos, V. (1990) *J. Biol. Chem* 265:15015-15022; Yanagibashi, K. et al. (1988) *Endocrinology* 123: 2075-2082; Besman, M. J. et al. (1989) *Proc. Natl. Acad. Sci. U.S.A.* 25 86:4897-4901; Papadopoulos, V. et al. (1991) *Endocrinology* 129:1481-1488).

Based on the amino acid sequence of the 18-kDa PBR, a three dimensional model was developed (Papadopoulos, V. (1996) In: *The Leydig Cell*. Payne, A.H. et al. (eds) Cache River Press, IL, pp. 596-628). This model was shown to accomodate a cholesterol molecule and function as a channel, supporting the role of PBR in cholesterol transport. 30 Recently we demonstrated the role of PBR in steroidogenesis by generating PBR negative cells by homologous recombination (Papadopoulos, V. et al. (1997) *J. Biol. Chem.* 272:32129-32135) that failed to produce steroids. However, addition of the hydrosoluble analogue of cholesterol, 22R-hydroxycholesterol, recovered steroid production by these

cells, indicating that the cholesterol transport mechanism was impaired. Further cholesterol transport experiments in bacteria expressing the 18-kDa PBR protein provided definitive evidence for a function as a cholesterol channel/transporter (Li and Papadopoulos, V. *et al.* (1998) *Endocrinology*).

5           We hypothesized that the peripheral-type benzodiazepine receptor is part of the changes in cellular and molecular functions that account for the increased aggressive behavior in cancer, and we chose to examine this hypothesis in human breast cancer. Breast cancer is the most common neoplasm and the leading cause of cancer-related deaths for women in most developing countries (Lippman, M. E. (1993) *Science* 259:631-  
10   632), affecting nearly 184,000 women, with over 46,000 deaths annually in the U.S. alone (American Cancer Society, 1996). Human breast cells are unlike brain and gonadal cells and cannot produce steroids, but like many other cells in the body, are able to metabolize steroids.

          Increased PBR expression correlates with increased aggressive behavior of tumor  
15   cells. Invasive tumors invade and grow locally but they do not metastasize. However, the aggressive tumors have the ability to invade and metastasize through the blood vessels to different places of the human body. Tumor metastasis into vital organs (such as lungs) is the most common cause of death.

          The correlation between high levels of expression of PBR and metastatic potential in  
20   for human breast cancer is shown in copending U.S. Application No. 09/047,652 filed March 25, 1998, in which Vassilios Papadopoulos of the instant application is a co-inventor. However, due to the involvement of PBR in cell proliferation, and the expression of PBR in all cells, it is likely that this correlation would exist for other solid tumors and cancers such as prostate cancer, colon cancer, brain tumors, and tumors in steroid producing tissues  
25   such as gonadal tumors, to name a few.

#### **Summary of the Invention**

          In one aspect, the present invention is directed to a method of combating cancer in a patient in need of such combating, wherein the cancer is caused by the deregulation of expression of proteins having a role in regulating tumor cells, which comprises administering  
30   an effective amount of Ginkgo biloba extracts or isolated Ginkgolide B to said patient.

          In another aspect, the present invention is directed to a method of combating the proliferation of cancer cells in a patient in need of such combating, wherein the proliferation is caused by the deregulation of expression of proteins having a role in regulating tumor

cells, which comprises administering an effective amount of Ginkgo biloba extracts or isolated Ginkgolide B to said patient.

In another aspect, the present invention is directed to a method of combating the proliferation of cancer cells in a patient in need of such combating, wherein the proliferation  
5 is caused by over-expression of proteins having a role in regulating tumor cells, which comprises administering an effective amount of Ginkgo biloba extracts or isolated Ginkgolide B to said patient.

In another aspect, the present invention is directed to a method of combating the proliferation of cancer cells having an aggressive phenotype in a patient in need of such  
10 combating, wherein the proliferation is caused by the over-expression of peripheral-type benzodiazepine receptor protein, which comprises administering an effective amount of Ginkgo biloba extracts or isolated Ginkgolide B to said patient.

In another aspect, the present invention is directed to a method of combating the proliferation of cancer cells, where the proliferation is caused by the over-expression of  
15 oncogenes, by decreasing the expression of said oncogenes in a patient in need of such combating, which comprises administering an effective amount of Ginkgo biloba extracts or isolated Ginkgolide B to said patient. A preferred method of the immediately foregoing method is where said oncogenes are one or more of APC, PE-1, RhoA and c-Jun.

In another aspect, the present invention is directed to a method of decreasing the  
20 expression of peripheral-type benzodiazepine receptor in cancer cells in a patient in need of such decreasing, wherein said cancer cells express an abnormal level of peripheral-type benzodiazepine receptor relative to normal cancer cells, which comprises administering an effective amount of Ginkgo biloba extracts or isolated Ginkgolide B to said patient. A preferred method of the immediately foregoing method is where the cancer cells are human  
25 breast cancer cells; human glioblastomas; human brain tumors; human astrocytomas; human colonic carcinoma; human colonic adenocarcinoma; human ovarian carcinomas; and human hepatocellular carcinoma.

In another aspect, the present invention is directed to a method of decreasing the expression of peripheral-type benzodiazepine receptor mRNA in cancer cells in a patient in  
30 need of such decreasing, which comprises administering an effective amount of Ginkgo biloba extracts or isolated Ginkgolide B to said patient.

In another aspect, the present invention is directed to a method of increasing the expression of c-Myc protooncogene in a patient in need of such increasing, which comprises



administering an effective amount of Ginkgo biloba extracts or isolated Ginkgolide B to said patient.

In another aspect, the present invention is directed to a method of decreasing the expression of cell cycle regulators prothymosin- $\alpha$ , CDK2, p55CDC, myeloblastin and p120  
5 proliferating-cell nuclear antigen (PCNA) in a patient in need of such decreasing, which comprises administering an effective amount of Ginkgo biloba extracts or isolated Ginkgolide B to said patient.

In another aspect, the present invention is directed to a method of decreasing the expression of intracellular signal transduction modulators NET1 and ERK2, in a patient in  
10 need of such decreasing, which comprises administering an effective amount of Ginkgo biloba extracts or isolated Ginkgolide B to said patient.

In another aspect, the present invention is directed to a method of decreasing the expression of apoptosis-related proteins Adenosine A2A Receptor, Flt3 ligand, Grb2, Clusterin, RXR- $\beta$ , Glutathione S-transferase P, N-Myc, TRADD, SGP-2 and NIP-1, in a  
15 patient in need of such decreasing, which comprises administering an effective amount of Ginkgo biloba extracts or isolated Ginkgolide B to said patient.

In another aspect, the present invention is directed to a method of decreasing the expression of transcription factors Id-2, ATF-4, ETR101 and ETR-103 in a patient in need of  
20 such decreasing, which comprises administering an effective amount of Ginkgo biloba extracts or isolated Ginkgolide B to said patient.

In another aspect, the present invention is directed to a method of decreasing the expression of growth factors macrophage colony-stimulating factor-1, heparin-binding EGF-like growth factor, hepatocyte growth factor-like protein and inhibin  $\alpha$ , in a patient in need of  
25 such decreasing, which comprises administering an effective amount of Ginkgo biloba extracts or isolated Ginkgolide B to said patient.

In another aspect, the present invention is directed to a method of decreasing the expression of cell adhesion molecules CD19 B-lymphocyte antigen, L1CAM,  $\beta$ -catenin, integrin subunits  $\alpha$ 3,  $\alpha$ 4,  $\alpha$ 6,  $\beta$ 5, and  $\alpha$ M, in a patient in need of such decreasing, which  
30 comprises administering an effective amount of Ginkgo biloba extracts or isolated Ginkgolide B to said patient.

In another aspect, the present invention is directed to a method of decreasing the expression of genes APC, PE-1, RhoA, c-Jun, prothymosin- $\alpha$ , CDK2, p55CDC, myeloblastin, p120 proliferating-cell nuclear antigen (PCNA), NET1, ERK2, Adenosine A2A

Receptor, Flt3 ligand, Grb2, Clusterin, RXR- $\beta$ , Glutathione S-transferase P, N-Myc, TRADD, SGP-2, NIP-1, Id-2, ATF-4, ETR-101, ETR-103, macrophage colony-stimulating factor-1, heparin-binding EGF-like growth factor, hepatocyte growth factor-like protein, inhibin  $\alpha$ , CD19 B-lymphocyte antigen, L1CAM,  $\beta$ -catenin, and integrin subunits  $\alpha$ 3,  $\alpha$ 4,  $\alpha$ 6,  $\beta$ 5, and  $\alpha$ M, in a patient in need of such decreasing, which comprises administering an effective amount of Ginkgo biloba extracts or isolated Ginkgolide B to said patient.

In another aspect, the present invention is directed to a pharmaceutical composition comprising an effective amount of Ginkgo biloba extracts or isolated Ginkgolide B for combating cancer and a pharmaceutically acceptable carrier or diluent.

Of all of the foregoing methods and compositions of the present invention, a preferred embodiment of each is where the Ginkgo biloba extracts is EGB 761®.

Further, of all the foregoing methods and compositions of the present invention, it is preferred that Ginkgolide B is used.

#### **Brief Description of the Drawings**

The file of this patent contains at least one drawing executed in color (Figure 11). This photograph is retained by the International Bureau as part of the record copy.

**Figure 1.** Effect of various concentrations of EGB 761® on MDA-231 PBR ligand binding capacity. MDA-231 cells were cultured as described under Materials and Methods. Cells were treated with the indicated concentrations of the injectable form of EGB 761®. At the indicated time periods cells were collected and PBR ligand binding characteristics were determined by Scatchard analysis. Data points represent the mean  $\pm$  S.D. of three independent experiments carried out in triplicate.

**Figure 2.** Effect of various concentrations of GKB on MDA-231 PBR ligand binding capacity. MDA-231 cells were cultured as described under Materials and Methods. Cells were treated with the indicated concentrations of GKB for 48 hours. At the indicated time periods cells were then collected and PBR ligand binding characteristics were determined by Scatchard analysis. Data points represent the mean  $\pm$  S.D. of two independent experiments carried out in triplicate.

**Figure 3.** Effect of EGB 761® and GKB on PBR mRNA levels in MDA-231 cells. Cells were treated for 48 hours without or with either 20 (EGb-20) or 200 (EGb-200)  $\mu$ g/ml EGB 761® or 2 (GKB-2) or 20 (GKB-20)  $\mu$ g/ml GKB. At the end of the incubation total RNA was isolated and loaded onto a 1% formaldehyde gel at a concentration of 10 $\mu$ g/lane. Northern blots were incubated with  $^{32}$ P-labeled hPBR probe and exposed to XOMAT Kodak film. Top,

autoradiogram of the blot. PBR migrates at 0.9Kb. Bottom, relative intensity of the PBR mRNA/28S ribosomal RNA visualized by ethidium bromide staining. The autoradiogram and PBR mRNA quantitation represent one out of two independent experiments.

**Figure 4.** Effect of EGB 761® on MDA-231 cell proliferation. MDA-231 cells grown in 96-well plates were washed with PBS and cultured in media supplemented with 10% FBS in the presence or absence of the indicated concentrations of EGB 761®. 4h prior to the end of incubation, bromodeoxyuridine (BrdU) was added to each well. Incorporation of BrdU was measured at 450nm (reference=700nm). Data points represent the mean  $\pm$  S.D. of four independent experiments carried out in quadruplicate. One-way ANOVA indicates that MDA-231 cell proliferation was significantly altered by treatment with EGB 761® at 48, 72 and 96 h timepoints ( $P<0.0001$ ).

**Figure 5 Right, Middle, Left.** Recovery of MDA-231 cell proliferation upon removal of EGB 761®. MDA-231 cells grown in 96-well plates were washed with PBS and cultured in media supplemented with 10% FBS in the presence or absence of 2 (Left), 20 (Middle) or 200 (Right)  $\mu$ g/ml EGB 761® for 48 h. At the end of the treatment the cells were washed and incubated in EGB 761®-free media for 48 h. 4h prior to the end of incubation, bromodeoxyuridine (BrdU) was added to each well. Incorporation of BrdU was measured at 450nm (reference=700nm). Data points represent the mean  $\pm$  S.D. of two independent experiments carried out in quadruplicate.

**Figure 6.** Effect of GKB on MDA-231 cell proliferation. MDA-231 cells grown in 96-well plates were washed with PBS and cultured in media supplemented with 10% FBS in the presence or absence of either 2  $\mu$ g/ml or 20  $\mu$ g/ml GKB for 48 hours. 4h prior to the end of incubation, bromodeoxyuridine (BrdU) was added to each well. Incorporation of BrdU was measured at 450nm (reference=700nm). Data points represent the mean  $\pm$  S.D. of two independent experiments carried out in quadruplicate. One-way ANOVA indicates that MDA-231 cell proliferation was significantly altered by treatment with GKB ( $P<0.0001$ ).

**Figure 7.** Effect of EGB 761® and GKB on MCF-7 cell proliferation. MCF-7 cells were grown in 96-well plates as described in Figure 4 for the MDA-231 cells. MCF-7 cells were treated for 48 hours without or with either 20 (EGB-20) or 200 (EGB-200)  $\mu$ g/ml EGB 761®, or 2 (GKB-2) or 20 (GKB-20)  $\mu$ g/ml GKB. 4h prior to the end of incubation, bromodeoxyuridine (BrdU) was added to each well. Incorporation of BrdU was measured at 450nm (reference=700nm). Data points represent the mean  $\pm$  S.D. of two independent experiments carried out in quadruplicate. One-way ANOVA indicates that MCF-7 cell

proliferation was altered to a lesser degree than the MDA-231 cell proliferation by treatment with EGB 761® or GKB.

**Figure 8.** Effect of EGB 761® on MDA-231 cell free radical production. MDA-231 cells were treated for 48 hours with 2 (Egb-2), 20 (Egb-20) or 200 (Egb-200) µg/ml of EGB 761®.

5 The cells were then washed and the levels of cellular oxidative stress were measured using the fluorescent probe DCF as described under Materials and Methods. Results shown are means ± S.D. (n=4). Statistical analysis indicated that the effect of EGB 761® was not significant.

**Figure 9.** Transcriptional response to EGB 761® suggests an effect on genes involved in cell proliferation. Results shown represent quantitative analysis of the Atlas human cDNA expression array containing 588 PCR-amplified cDNA fragments (Clontech Inc.). mRNAs were obtained from control or EGB 761® (20 µg/ml) treated, for 48 h, MDA-231 cells. For normalizing the mRNA abundance, the densitometric values obtained from image analysis were normalized using the housekeeping genes provided in the array. Only consistent  
10  
15 significant changes above 30% were considered.

**Figure 10.** Growth of MDA-231 xenografts in nude mice following either EGB 761® or GKB treatments. Animals were treated either orally with 50 mg/kg EGB 761® or ip with 1 mg/kg GKB once a day for a month starting with 100-150 mm<sup>3</sup> volume MDA-231 tumors. After the end of the treatment the animals were kept for 30 more days and then the animals were  
20 sacrificed on day 60. Data shown are means ± S.E.M. (n=10). Statistical analysis indicated that the effects of EGB 761® and GKB were significant compared to their respective controls ( $p<0.05$ ).

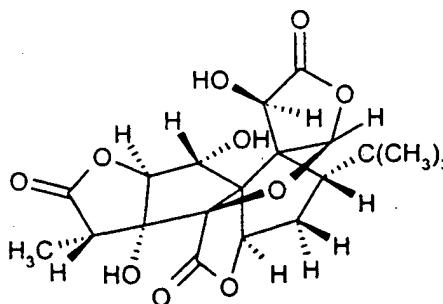
**Figure 11 A, B, C, D, E, F, G, H.** PBR expression in MDA-231 xenografts from control and EGB 761® or GKB treated animals. Formalin embedded sections of MDA-231 xenografts were immunostained with an anti-PBR antiserum at 1:500 dilution as described in the  
25 Materials and Methods section. MDA-231 tumors were obtained from animals treated with vehicle (A-D), 50 mg/kg EGB 761® per os (E, G), or 1 mg/kg GKB ip (F, H). A shows cells found in the middle of a tumor obtained from vehicle-treated animals where the nuclear localization of the PBR protein can be easily seen (see arrowheads). B shows the immunostaining seen in cells present in the edge of the tumor obtained from vehicle-treated animals. A higher magnification of the immunostaining seen in the cells present in the edge of the tumor obtained from vehicle-treated animals is shown in C. D represents a control  
30 treated with a non-specific antiserum. E shows the PBR immunostaining in cells found in the

middle of the tumor obtained from animals treated with EGB 761®. **F** shows the PBR immunostaining in cells found in the middle of the tumor obtained from animals treated with GKB. **G** shows the PBR immunostaining in cells found in the edge of the tumor obtained from animals treated with EGB 761®. **H** shows the PBR immunostaining in cells found in the middle of the tumor obtained from animals treated with GKB. Arrowheads indicate nuclei. Magnification is x75 for **A**, **B**, **C**, **E**, **F**, **G**, and **H** and x150 for **C**.

### Detailed Description

The term "ginkgo terpenoid" includes all of the naturally occurring terpenes which are derived from the gymnosperms tree *Ginkgo biloba* as well as synthetically produced ginkgo terpenoids and pharmaceutically active derivatives and salts thereof and mixtures thereof. Examples of ginkgo terpenoids include ginkgolides. Examples of ginkgo terpenoids are disclosed in *Ginkgolides, Chemistry, Biology, Pharmacology, and Clinical Perspectives*, J.R. Provs. Science Publishers, Edited by P. Braguet (1988); F.V. DeFeudis, *Ginkgo Biloba Extract (EGB 761®)*; *Pharmacological Activities and Clinical Applications*, Elsevier, Chapter II (1991).

The term "ginkgolide" as used herein include the various ginkgolides disclosed in the books cited above as well as non-toxic pharmaceutically active derivatives thereof. Examples of ginkgolide derivatives include tetrahydro derivatives, acetyl derivatives, and alkyl esters such as the monoacetate derivatives and triacetate derivatives disclosed in Okabe, et al., J. Chem. Soc. (c), pp. 2201-2206 (1967). Ginkgolide B has the following structure and as used herein, refers to isolated ginkgolide B:



The term "Ginkgo biloba extract" as used herein includes a collection of natural molecules, including terpenoids, derived from the leaves of the Ginkgo biloba tree. Preferably, the extract is the specific formulation of *Ginkgo biloba* extract known as EGB 761®.

The term "combating" as used herein means preventing, inhibiting and or decreasing whatever the word "combating" acts upon, e.g., combating cancer cell proliferation means that the cancers cells are prevented and inhibited from proliferating further and or the degree or rate of proliferation is decreased.

- 5           The level of expression of PBR, for the purposes of diagnosis or prognosis of a cancer or tumor, can be detected at several levels. Using standard methodology well known in the art, assays for the detection and quantitation of PBR RNA can be designed, and include northern hybridization assays, in situ hybridization assays, and PCR assays, among others. See e.g., Maniatis, Fritsch and Sambrook, *Molecular Cloning; A Laboratory Manual* 10 (1982) or *DNA Cloning*, Volumes I and II (D. N. Glover ed. 1985), or *Current Protocols in Molecular Biology*, Ausubel, F. M. *et al.* (Eds), Wiley & Sons, Inc. for the general description of methods for nucleic acid hybridization. Polynucleotide probes for the detection of PBR RNA can be designed from the sequence available at accession number L21950 for the human PBR sequence (Riond, J. *et al.* (1991) *Eur. J. Biochem.* **195**:305-311; Chang, Y. J. 15 *et al.* (1992) *DNA and Cell Biol.* **11**:471-480). The sequence of PBR from other sources such as bovine (Parola, A. L. *et al.* (1991) *J. Biol. Chem* **266**:14082-14087) and mouse (Garnier, M. *et al.* (1994) *Mol Phar.* **45**:201-211) are also known.

- The complete sequence of the PBR, normal or mutant, can be used for a probe to detect RNA expression. Alternatively, a portion or portions of the sequence can be used.
- 20       Methods for designing probes are known in the art. Polynucleotide sequences are preferably homologous to or complementary to a region of the PBR gene, preferably, the sequence of the region from which the polynucleotide is derive is homologous to or complementary to a sequence which is unique to the PBR gene. Whether or not a sequence is unique to the PBR gene can be determined by techniques known to those of 25 skill in the art. For example, the sequence can be compared to sequences in databanks, e.g., GenBank. Regions from which typical DNA sequences may be derived include but are not limited to, for example, regions encoding specific epitopes, as well as non-transcribed and/or non-translated regions.

- PBR ligands or anti-PBR antibodies, or fragments of ligand and antibodies capable 30 of detecting PBR may be labeled using any of a variety of labels and methods of labeling for use in diagnosis and prognosis of disease, such as breast cancer, particularly for assays such as Positron Emission Tomography and magnetic resonance imaging (Leong, D. *et al.* (1996) *Alcohol Clin. Exp. Res.* **20**: 601-605). Examples of types of labels which can be used

invention include but are not limited to enzyme labels, radioisotopic labels, non-radioactive isotopic labels and chemiluminescent labels.

Examples of suitable enzyme labels include malate dehydrogenase, staphylococcal nuclease, delta-5-steroid isomerase, yeast-alcohol dehydrogenase, alpha-glycerol phosphate dehydrogenase, triose phosphate isomerase, peroxidase, alkaline phosphatase, asparaginase, glucose oxidase, beta-galactosidase, ribonuclease, urease, catalase, glucose-6-phosphate dehydrogenase, glucoamylase, acetylcholine esterase, etc.

Examples of suitable radioisotopic labels include  $^3\text{H}$ ,  $^{111}\text{In}$ ,  $^{125}\text{I}$ ,  $^{32}\text{P}$ ,  $^{35}\text{S}$ ,  $^{14}\text{C}$ ,  $^{57}\text{Co}$ ,  $^{59}\text{Fe}$ ,  $^{75}\text{Se}$ ,  $^{152}\text{Eu}$ ,  $^{90}\text{Y}$ ,  $^{67}\text{Cu}$ ,  $^{21}\text{Cl}$ ,  $^{211}\text{At}$ ,  $^{212}\text{Pb}$ ,  $^{47}\text{Sc}$ ,  $^{109}\text{Pd}$ ,  $^{11}\text{C}$ ,  $^{19}\text{F}$ ,  $^{123}\text{I}$ , etc.

Examples of suitable non-radioactive isotopic labels include  $^{157}\text{Gd}$ ,  $^{55}\text{Mn}$ ,  $^{162}\text{Dy}$ ,  $^{52}\text{Tr}$ ,  $^{46}\text{Fe}$ , etc.

Examples of suitable fluorescent labels include a  $^{152}\text{Eu}$  label, a fluorescein label, an isothiocyanate label, a rhodamine label, a phycoerythrin label, a phycodyanin label, an allophycocyanin label, a fluorescamine label, etc.

Examples of chemiluminescent labels include a luminal label, an isoluminal label, an aromatic acridinium ester label, an imidazole label, an acridinium salt label, an oxalate ester label, a luciferin label, a luciferase label, etc.

We examined herein the effect of Ginkgo biloba extracts, more specifically EGB 761® and GKB on PBR expression and cell proliferation, particularly in human breast cancer cells. We used the highly aggressive cell line MDA-231, which expresses over 60-fold higher levels of PBR ligand binding and mRNA relative to the non-aggressive cell line MCF-7. EGB 761® and GKB decreased in a time- and dose-dependent manner PBR expression and cell proliferation in MDA-231 cells whereas EGB 761® and GKB did not affect the MCF-7 cell proliferation to the same degree. This effect was reversible and it was not due to the antioxidant properties of the compounds tested.

The determination of elevated levels of PBR is done relative to a sample with no detectable tumor. This may be from the same patient or a different patient. For example, a first sample may be collected immediately following surgical removal of a solid tumor. Subsequent samples may be taken to monitor recurrence of tumor growth and/or tumor cell proliferation. Additionally, other standards may include cells of varying aggressive phenotype such that an increase or decrease in aggressive phenotype can be accessed.

The distinct sub-cellular localization of PBR in the cytoplasm of epithelial cells of normal breast ducts and the absence of staining in the nucleus and the perinuclear area of

the aggressive tumor cells provides a simple method for diagnosing the aggressive phenotype of a tumor cell. Immunostaining using labeled PBR ligand or labeled PBR antibody or fragment of ligand or antibody capable of binding to PBR and determining the sub-cellular location of PBR in the cellular samples provides yet another diagnostic assay of  
5 the present invention. In addition, antiserum which recognizes PBR can also be used along with a secondary antibody reactive with the primary antibody. Immunostaining assays are well known in the art, and are additionally described in the Examples below with respect to breast cancer cells and biopsies.

An increase in the level of PBR is determined when the level of PBR in a tumor cell  
10 is about 2-3 times the level of PBR in the normal cell, up to about 10-100 times the amount of PBR in a normal cell.

**Cell Culture and Treatments.** Human breast cancer cell lines (MCF-7 and MDA-231) were obtained from the Lombardi Cancer Center, Georgetown University Medical Center. Cell lines were cultured on polystyrene culture dishes (Corning) and grown in Dulbecco's  
15 modified Eagle medium (DMEM) supplemented with 10% fetal bovine serum (FBS). The injectable form (IPS200) of the standardized *Ginkgo biloba* extract EGB 761® was used. This injectable form is devoid of procyanidins, which are known to interact with proteins *in vitro* (Defeuder, 1998 Ullstein Medical). The injectable form of EGB 761® and GKB (BN 52021) isolated from EGB 761® were provided by the Institut Henri Beaufour-IPSEN (Paris,  
20 France).

**Radioligand Binding Assays.** Cells were scraped from 150mm culture dishes into 5ml phosphate buffered saline (PBS), dispersed by trituration, and centrifuged at 500xg for 15min. Cell pellets were re-suspended in PBS and assayed for protein concentration. [<sup>3</sup>H]PK 11195 binding studies on 50μg of protein from cell suspensions were performed as  
25 previously described (Papadopoulos, V. et al., 1990, J. Biol. Chem. 265: 3772-3779; Hardwick, M. et al., 1999, Cancer Research, 59: 631-632) the contents of which are incorporated herein by reference. [N-methyl-<sup>3</sup>H]PK 11195 (1-(2-chlorophenyl)-N-methyl-N-(1-methyl-propyl)-3-isoquinolinecarboxamide; sp. Act. 83.50 (Ci/mmol), was obtained from Du Pont-New England Nuclear (Wilmington, DE) and PK 11195 was obtained from  
30 Research Biochemicals Incorporated (Natick, MA). Scatchard plots were analyzed by the LIGAND program (Munson, PJ, and Robbard, D. 1980, Anal. Biochem., 107: 220-239) (BIOSOFT, Ferguson, MO).



**Protein Measurement.** Protein levels were measured by the Bradford method (Bradford, MM, 1976, Anal. Biochem., 72: 248-254) using the Bio-Rad Protein Assay kit (Bio-Rad Laboratories, Hercules, CA) with bovine serum albumin as a standard.

**RNA (Northern) Analysis.** PBR mRNA expression in MDA-231 cells treated with the various compounds was examined by Northern Blot analysis as we previously described (Hardwick, M., et al., 1999, Cancer Research, 59: 831-842). In brief, total cellular RNA was isolated using the RNeasy B reagent (TEL-TEST, Inc., Friendswood, TX) and chloroform. 20µg of total RNA from each cell line were run on 1% agarose gels and transferred overnight to nylon membranes (S&S Nytran, Schleicher & Schuell, Keene, NH) (21). A 0.2 kb human PBR (hPBR) cDNA fragment (derived from the pCMV5-PBR plasmid vector containing the full length hPBR kindly given by Dr. Jerome Strauss, University of Pennsylvania, PA) was radiolabeled with [ $\alpha$ -<sup>32</sup>P]dCTP using a random primers DNA labeling system (Life Technologies, Gaithersburg, MD). The hybridization conditions were as we previously described (Hardwick, M., et al., 1999, Cancer Research, 59: 831-842). Autoradiography was performed by exposing the blots to X-OMAT AR film (Kodak, Rochester, NY) at -70°C for 4-48hr. Quantification of PBR mRNA was carried out using the SigmaGel software (Jandel Scientific, San Rafael, CA).

**Nucleic Acid Arrays.** We used the Atlas human cDNA expression array I from Clontech (Palo Alto, CA). This array contains 588 human PCR-amplified cDNA fragments of 200-500 bp long immobilized on a positively charged nylon membrane. MDA-231 cells were treated with and without 20 µg/ml EGB 761® for 48 hours. Poly A+ RNA was isolated from control and EGB 761®-treated cells. <sup>32</sup>P-labeled cDNA probes were generated from each poly A+RNA and hybridized to the Atlas array according to the manufacturer's recommendations. Autoradiography was performed by exposing the blots to X-OMAT AR film (Kodak, Rochester, NY) at -70°C for 4-96 hr. Quantification of the hybridization seen was carried out using the SigmaGel software (Jandel Scientific, San Rafael, CA). Multiple exposures were used in order to detect genes expressed at low levels. The three internal controls, ubiquitin, G3PDH and  $\beta$ -actin were used to compare the relative expression levels of the detected gene products in the control and EGB 761®-treated cells. Experimental variations were corrected using the ratios of gene expression versus the internal controls. The effect of the EGB 761® treatment on each gene product is expressed as % of control (untreated) cells. The results presented herein show genes affected consistently, at a level above 30% of control, by the EGB 761® treatment.

**BrdU Cell Proliferation Assays.** MDA-231 cells were plated on 96-well plates (Corning, Corning, NJ) at a concentration of approximately 10,000 cells/well (24h incubation) or approximately 5,000 cells/well (48h incubation) in DMEM supplemented with 0.1% FBS. The cells were then incubated in 10% FBS with various concentrations of EGB 761® or GKB for the indicated time periods. Differences in cell proliferation were analyzed by measuring the amount of 5-bromo-2'-deoxyuridine (BrdU) incorporation determined by the BrdU ELISA (Boehringer Mannheim, Indianapolis, IN). Incorporation of BrdU was measured at 450 nm (reference at 690 nm).

**Analysis of oxidative stress.** Levels of cellular oxidative stress were measured using the fluorescent probe 2,7-dichlorofluorescein diacetate (2,7-DCF; Molecular Probes, Inc., Eugene, OR) as described in Goodman, Y. and Mattson, M.P., *Exp. Neurol.*, **128**: 1-12, 1994. In brief, cells were cultured in 96-well plates and treated for 48 hours with the indicated concentrations of EGB 761®. At the end of the treatment the cells were washed and incubated in the presence of 50  $\mu$ M 2,7-DCF in PBS. Fluorescence was then quantified using the Victor<sup>2</sup> quantitative detection fluorometer (EGG-Wallac, Inc., Gaithersburg, MD).

**Biological Evaluation *In Vivo*.** The MDA-231 human breast cancer (estrogen insensitive) xenograft model was used for *in vivo* screening of EGB 761® and GKB. Based on the *in vitro* data and previously published *in vivo* data (23) the doses used were 50 mg/kg for the EGB 761® and 1 mg/kg for the GKB. Female athymic nude mice (NCI/Charles River, Frederick, MD) are injected subcutaneously with  $8 \times 10^5$  MDA-231 tumor cells and tumors are allowed to form to a volume of ~100 to 150 mm<sup>3</sup>. At this time, groups of 10 animals per compound were injected either orally for the EGB 761® or intraperitoneally for the GKB once a day for a month. Twice weekly tumor sizes and body weights were recorded for all animals for the 30 days of treatment as well for 30 days after the end of the treatment. At that time the animals were sacrificed and the tumors were removed and processed for immunohistochemistry. Animal care was in accordance with institutional guidelines.

**Immunocytochemistry of MDA-231 tumors.** MDA-231 tumors removed from the mice treated with or without EGB 761® or GKB were fixed in 10% buffered formalin. Tumors were sectioned and then placed on glass slides and processed as we previously described (19). For immunohistochemistry with anti-PBR primary antibodies, tissue sections were treated with a 30% H<sub>2</sub>O<sub>2</sub>/methanol mixture (1:9 ratio) for 5 min at room temperature to neutralize endogenous peroxidase activity and then washed well with PBS. Primary antibody in 10% calf serum in PBS was added to sections at a concentration of 1:500 at RT

for 1h. Secondary antibody reactions were performed using horseradish peroxidase-coupled goat anti-rabbit secondary antibody diluted 1:500 in PBS supplemented with 10% calf serum. After washing the slides three times in PBS for 2 min each, fresh H<sub>2</sub>O<sub>2</sub> diluted 1:1,000 with 3-amino-9-ethyl carbazole (AEC) was added and slides were incubated for 1h at 37°C. The slides were then rinsed in distilled H<sub>2</sub>O before mounting with Crystal/Mount.

**Statistical Analysis.** Comparison of multiple means was performed with InStat's one-way analysis of variance (ANOVA) (GraphPad Inc., San Diego, CA). All *F* statistics and *P* values for one-way ANOVAs are provided in the text. Comparison of individual drug treatments to the control treatments was performed with unpaired *t*-test. All *p* values for unpaired *t*-tests are provided in the text.

## RESULTS

**EGB 761® and GKB reduce the PBR Ligand Binding Capacity of the MDA-231 Human Breast Cancer Cells.** Figure 1 shows that increasing concentrations of the injectable form of EGB 761® inhibit in time-dependent manner the PBR ligand binding capacity (B<sub>max</sub>), determined using saturation isotherms with the radiolabeled ligand PK 11195 followed by Scatchard analysis of the data. Similar results were obtained using isolated GKB (Figure 2). Interestingly, EGB 761® and GKB decreased PBR levels by 66% of control values. No significant effects on the receptor affinity (K<sub>d</sub>) could be seen ( $5.8 \pm 1.4$  pmol/mg protein, n=12).

**EGB 761® and GKB reduce the PBR mRNA Expression in MDA-231 Human Breast Cancer Cells.** RNA (Northern) blot analysis was performed in order to determine if the differences seen in PBR ligand binding between the control and the EGB 761®- or GKB-treated cells reflect an effect on PBR mRNA expression. As shown in Fig. 3, both EGB 761® and GKB reduced PBR mRNA levels. This result fits with the results presented above on the PBR ligand binding expression.

**EGB 761® and GKB Inhibit MDA-231 Cell Proliferation.** Using the Bromodeoxyuridine (BrdU) Cell Proliferation ELISA (Boehringer-Mannheim, Indianapolis, IN), we examined the effect of increasing concentrations of EGB 761® on MDA-231 cell proliferation. Fig. 4 shows that EGB-761 inhibits in a concentration- and time-dependent manner the MDA-231 cell proliferation. This effect of EGB 761® was reversible, even for the highest concentration of EGB 761® used. (Fig. 5). Incubation of MDA-231 cells for 48 hours with EGB 761®, followed by washing and incubation for another 48 hours in EGB 761®-free medium,

resulted in the recovery of the MDA-231 proliferative activity. Increasing concentrations of GKB also inhibited the MDA-231 cell proliferation after 48 hours treatment (Fig. 6).

**EGB 761® and GKB Do Not Have The Same Level of Inhibition Against MCF-7 Cell Proliferation as They Do Against MDA-231 Cell Proliferation.**

5 Compared to the MDA-231 cells, the non-aggressive MCF-7 cells contain extremely low (<60 fold) PBR levels, as determined by both ligand binding studies and mRNA analyses, we examined whether EGB 761® and GKB affect the MCF-7 cell proliferation rate to the same degree as it affects MDA-231 cell proliferation rate. Fig. 7 clearly shows that neither EGB 761® nor GKB affect MCF-7 cell proliferation to the same degree as they affect the MDA-231 cell proliferation.

10 **EGB 761® Does Not Affect MDA-231 Free Radical Levels.** Recent *in vivo* and *in vitro* studies demonstrated that the terpene constituents of EGB 761®, including GKB, have anti-oxidant properties. In order to determine whether the anti-proliferative effect of EGB 761® was due to its anti-oxidant properties, we determined the free radical levels in MDA-231 cells treated with and without increasing concentrations of EGB 761® (Fig. 8). Although a  
15 20% decrease in free radical levels could be seen, this effect was neither statistically significant nor dose-dependent, indicating that the effect seen was not due to an EGB 761®-induced decrease of free radical levels in the cells.

**EGB 761® Regulates the MDA-231 Transcriptional Program Related to Cell Proliferation.** The results presented above indicate that EGB 761® and GKB inhibit PBR  
20 expression and cell proliferation in the PBR-rich and highly aggressive MDA-231 breast cancer cells. The non-aggressive MCF-7 cells, which contain extremely low PBR levels, did not respond to EGB 761® treatment to the same degree as that of the MDA-231 breast cancer cells. In order to determine whether the effect of EGB 761® (20 µg/ml for 48 hours) on MDA-231 cells was specific for PBR or whether other genes involved in cell proliferation  
25 were affected by the treatment, we used a cDNA array representing 588 distinct human genes. As noted under Materials and Methods, experimental variations were corrected using the ratios of gene expression versus the internal controls. The effect of the EGB 761® treatment on each gene product is expressed as % of control (untreated) cells. Only consistent changes above 30% of control values are presented in Fig. 9 and Table I.

30 **Biological Evaluation of the Effect of EGB 761® and GKB *in Vivo*** In order to assess the effect of EGB 761® and GKB on tumor cell proliferation and PBR expression in an *in vivo* setting we used the mammary fat pad xenograft implantation model (See Medina, D., J.

Mamm. Gland. Biol. Neopl. 1:5-19, 1996). Fig. 10 shows that 30 days treatment with either 50 mg/kg EGB 761® or with 1 mg/kg GKB resulted in a 35% ( $p=0.037$ ) and 32% ( $p=0.043$ ) decrease in tumor size, measured a month after the end of the treatment, respectively. These treatments did not affect the animal body weight (data not shown). Considering these

5 *in vitro* data on the effect of EGB 761® and GKB on PBR expression in MDA-231 cells, we examined whether EGB 761® and GKB also decreased PBR expression in the MDA-231 xenografts. Fig. 11(A-D) shows horseradish peroxidase (HRP) staining of the PBR antiserum used to detect the 18,000 molecular weight protein in MDA-231 xenografts from vehicle-treated animals. The hematoxylin counterstain was omitted in order to distinguish

10 the nuclear localization of PBR (19) in the tumors. Fig. 11A shows the middle of a tumor where the nuclear localization of the 18,000 PBR protein can be easily seen (see arrowheads). Fig 11B shows the immunostaining seen in the edge of the tumor obtained from vehicle-treated animals. A higher magnification of the immunostaining seen in the edge of the tumor obtained from vehicle-treated animals is shown in Fig. 11C, and a control treated

15 with a non-specific antiserum is shown in Fig. 11D. Treatment with either EGB 761® (Fig. 11E) or GKB (Fig. 11F) reduced the nuclear PBR expression present in cells found in the middle of the tumor. Interestingly, treatment with either EGB 761® (Fig. 11G) or GKB (Fig. 11H) also eliminated the nuclear PBR expression present in the cells at the edge of the tumors. However, in the later case cytosolic immunostaining could be seen (Figs. 11G and

20 H). These data were replicated in sections taken from xenografts grown in three separate animals.

It is of interest to note that even in the presence of high concentrations of either EGB 761® or GKB, PBR levels and rates of cell proliferation could not be reduced below 30% of normal values. This suggests that there is a minimum of PBR required to maintain

25 membrane integrity and cell function. It should be also noted that even at the highest concentrations used, neither EGB 761® nor GKB were toxic for the cells, because cell proliferation recovered upon removal of the compounds. These data suggest that these compounds are cytostatic and not cytotoxic. Additional cytotoxicity assays indicated that under the same conditions neither EGB 761® nor GKB induced any significant cell death.

30 The absence of any significant decrease in the amount of reactive oxygen species produced in the MDA-231 cells by EGB 761® or GKB suggests that their anti-oxidant properties were not responsible for decreasing PBR expression and cell proliferation in the

MDA-231 cells. These results indicate that these compounds may regulate PBR gene transcription either directly or indirectly.

The finding that EGB 761® and GKB decreased PBR expression and cell proliferation in the highly aggressive, nuclear PBR-expressing MDA-231 cells, but did not affect the non-aggressive MCF-7 cells, which have extremely low PBR levels and are devoid of nuclear PBR, to the same degree as they did for MDA-231 cells, provides an additional support to the hypothesis that the presence of PBR may be a determinant factor for the aggressive phenotype of breast tumor cells. Moreover, this observation further demonstrates the specificity of the effect of EGB 761® and GKB on targeting the regulation of PBR expression. This later finding brought us to two key questions raised by the current study: is the expression of other genes regulated by EGB 761®? is there a transcriptional program activated or inhibited by EGB 761®, where PBR is at the origin or a part of a cascade of events, responsible for altering the proliferation rate of MDA-231 cells? To address these questions, we utilized the Atlas Human cDNA Expression Array. As indicated in Table I, treatment of MDA-231 cells with the EGB 761® extract induced alterations in the transcriptional expression of 36 out of 588 genes examined. Not surprisingly, the vast majority of the affected genes have close ties to either cell proliferation, differentiation, or apoptosis. Perhaps the most telling of the effects EGB 761® has on the MDA-231 cell line is the down-regulation of the p120 proliferation-cell nuclear antigen. p120 is used as a prognostic indicator in breast cancer patients and prostate adenocarcinomas (Perlaky, L., et al., *Cancer Res.*, 52: 428-436, 1992; Zhuang, S.H. et al., *Endocrinology*, 139: 1197-1207, 1998). More importantly, however, p120 is an immunocytochemical marker of proliferating cells. Down-regulation of this proliferation marker by 68% thus confirms our data demonstrating that EGB 761® inhibits MDA-231 cell proliferation.

Using a human cDNA expression array we examined the effect of the EGB 761 treatment on the expression of 588 genes in MDA-231 cells. We found that the treatment increased the expression of the c-Myc protooncogene and decreased the expression of 35 gene products, including oncogenes (AP-1, PE-1, RhoA, n-Myc), cell cycle regulators (CDK2, p55CDC, PCNA p120), signal transduction modulators (NET1, ERK2), apoptosis-related products (SGP-2, NIP1) receptors (A2A, RXR-beta, Grb2), transcription factors (Id-2, ATF-4, ETR101, ETR-103), growth factors (HB-EGF, HGF-like), and cell adhesion molecules (CD19, L1CAM, integrins  $\alpha 3$ ,  $\alpha 4$ ,  $\alpha 6$ ,  $\beta 5$ , Mac-1,  $\beta$ -catenin) which are directly involved in various pathways regulating cell proliferation. Considering that the compounds

tested were effective only on the MDA-231-cells, which express high levels of PBR, these data suggest that the expression of nuclear PBR may be a determining factor for a tumor cell to acquire an aggressive and invasive phenotype.

5

**Table I:** Effect of EGB 761® on MDA-231 gene expression examined using the Atlas human cDNA expression array as described under Materials and Methods.

Name	% Change	Function	References
<b><u>Oncogenes and Tumor Suppressers</u></b>			
c-Myc	+75%	-basic helix-loop-helix-leucine zipper transcription factor -Myc/Max heterodimers induce cell-cycle progression, apoptosis, and malignant transformation	(37)
c-Jun	-78%	-part of the AP-1 transcription factor that regulates genes involved in cell proliferation	(38)
RhoA	-93%	-GTP-binding protein that is an important regulator of cell proliferation -RhoA inactivation inhibits HL60 cell proliferation	(39) (40)
APC	-59%	-APC mutations are associated with both hereditary and sporadic colorectal cancers -a negative post-translational regulator of $\beta$ -catenin	(41) (42)
PE-1	-42%	-transcription factor	(43)
<b><u>Cell Cycle Control Proteins</u></b>			
Prothymosin- $\alpha$	-79%	-acidic nuclear protein that is upregulated in proliferating thymocytes, lymphocytes from leukemia patients, and in malignant breast lesions	(44)
Myeloblastin	-66%	-a serine protease involved in leukemia cell differentiation	(45)
p55CDC	-63%	-similar to mitosis regulators CDC4 and CDC20 -expression positively correlated with cell proliferation status	(46)
p120 Proliferating-cell Nuclear Antigen	-68%	-nucleolar protein expressed in proliferating cells -a prognostic indicator for breast cancer patients and prostate adenocarcinoma	(47) (48)
CDK2	-83%	-cyclin-dependent tyrosine kinase involved in progression through the cell cycle -cyclin E/Cdk2 inactivates the retinoblastoma tumor suppressor to allow the cell to progress to S phase -Vitamin D inhibition of LNCaP cell proliferation coincided with a reduction in Cdk2 activity	(49) (50)
<b><u>Intracellular Transducers</u></b>			
NET1	-55%	-RhoA-specific guanine exchange factor -NIH3T3-transforming protein	(51)
ERK2	-46%	-member of the extracellular signal-related protein kinase family -activated upon cell stimulation	(52)
<b><u>Apoptosis-Related Proteins</u></b>			
Adenosine A2A Receptor	-40%	-G protein-coupled receptor involved in the cAMP signaling pathway	(53)
Flt3 ligand	-58%	-ligand for the Flt3 cytokine receptor tyrosine kinase -induces proliferation of leukemic myeloid cells	(54)
Grb2	-70%	-an adapter protein that links receptor tyrosine kinases to the Ras/MAPK signaling pathway via its SH2 domain	(55)
Clusterin	-54%	-a glycoprotein associated with cell adhesion and apoptosis -increased expression is linked to Alzheimer's disease	(56, 57) (58)
RXR- $\beta$	-55%	-retinoid-activated transcription factor -inhibition of chondrocyte proliferation by retinoic acid causes a reduction in RXR- $\beta$ mRNA expression	(59) (60)
Glutathione S-transferase P	-39%	-a multi-drug resistance gene that is overexpressed in various human tumors	(61, 62) (63)

N-Myc	-74%	-chemical inhibition of GST-P inhibits proliferation of Jurkat T cells -c-myc family member	(64)
TRADD	-51%	-associated with early-onset retinoblastoma -TNFR-associated death domain protein	(65)
NIP-1	-40%	-involved in TNFR-induced cell growth and differentiation -originally described as a yeast nuclear transport protein	(66)
		-part of the translation initiation factor 3 (eIF3) core complex <u>DNA-Binding/Transcription Factors</u>	(67)
Id-2	-65%	-a member of the Id helix-loop-helix family of transcriptional inhibitors	(68)
ATF4	-42%	-involved in proliferation of human pancreatic cancer cells -a member of the ATF/CREB family of transcription factors	(69)
ETR103	-65%	-regulates Ras-induced transformation of NIH3T3 cells -a macrophage-associated immediate early gene	(70)
ETR101	-60%	-a lymphocyte-associated immediate early gene	(71)

Name	% Change	Function	References
<u>Cell Surface Antigens and Adhesion Molecules</u>			
CD19 B-lymphocyte Antigen	-62%	-B-lymphocyte integral membrane protein -expression is down-regulated during retinoid-inhibition of lymphoblastoid B-cell proliferation	(72)
L1CAM	-72%	-neural cell adhesion molecule -increased L1CAM expression is associated with high-grade migration of glioma cells	(73)
$\beta$ -catenin	-58%	-involved in cadherin-mediated cell-cell interactions -interacts with the TCF/LEF transcription factors in the Wnt signaling pathway	(74)
Integrin Subunits			
$\alpha$ M	-41%	-mediates cellular adherence of human neutrophils with LFA-1 $\beta$ - $\alpha$ subunit of the elastase receptor	(75)
$\beta$ 5	-55%	- $\beta$ subunit of the vitronectin receptor (VR) -involved in cessation of oligodendrocyte proliferation	(76)
		-involved in murine retinal angiogenesis	(77)
$\alpha$ 4	-49%	-cross-linking $\alpha$ 4 integrins inhibits LB lymphoma cell proliferation -also involved in metastasis of melanoma and lymphoma cells	(78)
$\alpha$ 3	-77%	-a functionally perturbing $\alpha$ 3 integrin antibody inhibits human epithelial cell proliferation	(79)
$\alpha$ 6	-53%	-overexpression of $\alpha$ 6 integrin collaborates with ErbB2 to induce a more malignant phenotype in NIH3T3 cells	(80)
<u>Extracellular Signaling/Communication Proteins</u>			
Macrophage Colony-stimulating Factor-1 (CSF-1)	-31%	-regulates the proliferation, differentiation, and survival of monocytes, macrophages and their precursors -initiates a mitogenic signal that is required throughout G1 phase -CSF-1 stably transfected ovarian granulosa cells exhibit enhanced cell proliferation	(81)
Heparin-binding EGF-like Growth Factor (HB-EGF)	-62%	-overexpressed in numerous human glioma cell lines and in a majority of glioblastomas -stimulates human glioma cell proliferation	(82)
Hepatocyte Growth Factor-like Protein (HGFLP)	-81%	-a transmembrane protein tyrosine kinase found to be overexpressed in hepatoblastoma and in human primary liver carcinoma	(83)
Inhibin $\alpha$	-69%	-induces proliferation and migration of murine keratinocytes -a member of the inhibin family of heterodimeric growth factors -inhibin $\alpha$ is a marker of trophoblastic neoplasia and is highly expressed in virilizing adenomas	(84)
			(85)
			(86)
			(87)
			(88)
			(89)

Two members of the Myc family of transcription factors, c-Myc and n-Myc, were found to be grossly altered in this experiment. Expression of the proto-oncogene c-Myc was increased by 75% while expression of n-Myc was reduced by 74%. Both of these genes are overexpressed in several cancer types (Kim, C.J., et al., Virchows Arch., 434: 301-305,



1999; Dang, C.V., Mol. Cel. Biol., 19: 1-11, 1999) and are strongly correlated with tumor cell proliferation. Previous studies have shown that arrest of neuroblastoma cell growth by the tyrosine kinase inhibitor genistein is accompanied by down-regulation of n-Myc expression. This data fits extremely well with our cell proliferation and n-Myc data. Overexpression of c-Myc, however, is associated with a stimulation of cell proliferation in normal serum conditions. Overexpression of c-Myc induces cell death in the absence of serum or other survival factors. Taken together the data implies that deregulation of c-Myc expression requires the altered expression of other genes, as well.

In our microarray experiment, we discovered the deregulated expression of several other c-Myc-related genes. One such gene, *prothymosin  $\alpha$*  (*proT  $\alpha$* ), is induced by c-Myc. However, expression of *proT  $\alpha$*  is reduced by 79% rather than increased, as might be predicted by up-regulation of c-Myc. Further, expression of other c-Myc target genes such as *cdc25A*, *cyclin A*, and *cyclin E* are unaffected by treatment of MDA-231 cells with EGB 761®, suggesting either a treatment-specific or a cell line-specific short circuit in c-Myc-regulated gene transcription. Other data gathered from the microarray experiment further supports this hypothesis. c-Myc transcriptional regulation is under the control of APC and  $\beta$ -catenin. However, both of these genes are down-regulated by EGB 761® in MDA-231 cells (59 and 58%, respectively) while c-Myc is up-regulated. While some of this data appears to be contradictory, much of the published data on the role of c-Myc in cell proliferation, differentiation, and cell death also appears contradictory.

Similar to the altered regulation of c-Myc and Myc-related proteins by EGB 761®, the microarray experiment exposed disruption of several signaling molecules. EGB 761® treatment resulted in a 93% reduction in the expression of *RhoA*, a gene encoding a GTP-binding protein involved in numerous cellular phenomena, and a 55% reduction in *NET1* expression, a RhoA-specific guanine exchange factor. Interestingly, *RhoA* has been demonstrated to regulate cyclin E/Cdk2 activity in fibroblasts. Activation of cyclin E/Cdk2 complex is crucial to the progression of the cell cycle from G1 to S-phase. Regulation of cyclin E/Cdk2 activity has also been demonstrated by c-Myc. Although the significance of these two phenomena is not immediately obvious, it should be noted that expression of *Cdk2* is reduced by 83% by EGB 761®.

Other important signaling molecules are also down-regulated by EGB 761®. Expression of the adapter molecule *Grb2* is reduced by 70%. *Grb2* plays an important role in cellular signaling by physically linking signal transducers such as receptor tyrosine

kinases to the Ras/MAPK pathway. With regard to the MAPK pathway, expression of the MAP/ERK family member ERK2 is down-regulated by 46% and expression of the c-Jun transcription factor is reduced by 78%. Interestingly, it has been reported that EGB 761® is a suppressor of AP-1 transcription factor stimulated by phorbol esters. These data imply that  
5 the effects of EGB 761® on MDA-231 cell proliferation are accompanied by a broad reduction in mRNAs with functional relationships with one another.

Another interesting finding from the microarray experiment is the reduced expression of several integrins. The integrins are a large family of cell-cell and cell-extracellular matrix adhesion receptors that are composed of two transmembrane glycoprotein subunits, one  $\alpha$ -  
10 and one  $\beta$ -subunit. All of the integrins represented in Table I are involved in the regulation of cell proliferation in some way. For example, integrin  $\alpha$ M is part of the elastase receptor. ONO-5046, an elastase inhibitor, suppresses the proliferation of polyoma virus- and Kirsten sarcoma virus-transformed BALB/c3T3 cells and of Capan-1 pancreatic carcinoma cells. Moreover, integrin  $\alpha$ 4 has been implicated in the accumulation of distal metastases in  
15 melanoma, sarcoma, and lymphoma cell models. It is important to emphasize that integrins function as heterodimers of one  $\alpha$ - and one  $\beta$ -subunit. Reduced expression of either the  $\alpha$ - or the  $\beta$ -subunit is clearly important in the regulation of integrin receptor function.

The expression of some key growth factor genes, such as the hepatocyte growth factor-like and EGF-like growth factor, were also reduced by EGB 761®. It is possible that  
20 the EGB 761®-induced inhibition of cell proliferation may be due to the reduced expression of these growth factors that may act as autocrine regulators of cell growth.

EGB 761® and GKB are intended to be provided to recipient patient in an amount sufficient to combat cancer in said patient or in an amount sufficient to affect the expression (negatively or positively) of the gene products listed in Table 1, hereinabove. An amount is  
25 said to be sufficient to combat cancer if the dosage, route of administration, etc. of the EGB 761® and GKB are sufficient to influence such a response.

A composition is said to be "pharmacologically acceptable" if its administration can be tolerated by a recipient patient. Such an agent is said to be administered in a "therapeutically effective amount" if the amount administered is physiologically significant.  
30 An agent is physiologically significant if its presence results in a detectable change in the physiology of a recipient patient.

EGB 761® and GKB can be formulated according to known methods to prepare pharmaceutically useful compositions, whereby these materials, or their functional

derivatives are optionally combined in admixture with a pharmaceutically acceptable carrier vehicle. Suitable vehicles and their formulation, inclusive of other human proteins, e.g., human serum albumin, are described, for example, in Remington's Pharmaceutical Sciences (16<sup>TH</sup> ED., Osol, A. ed., Mack Easton PA. (1980)). In order to form a  
5 pharmaceutically acceptable composition suitable for effective administration, such compositions will contain an effective amount of the above-described compounds together with a suitable amount of carrier vehicle.

Additional pharmaceutical methods may be employed to control the duration of action. Control release preparations may be achieved through the use of polymers to  
10 complex or absorb the compounds. The controlled delivery may be exercised by selecting appropriate macromolecules (for example polyesters, polyamino acids, polyvinyl, pyrrolidone, ethylenevinylacetate, methylcellulose, carboxymethylcellulose, or protamine sulfate) and the concentration of macromolecules as well as the method of incorporation in order to control release. Another possible method to control the duration of action by  
15 controlled release preparations is to incorporate the compounds of the present invention into particles of a polymeric material such as polyesters, polyamino acids, hydrogels, poly(lactic acid) or ethylene vinylacetate copolymers. Alternatively, instead of incorporating these agents into polymeric particles, it is possible to entrap these materials in microcapsules prepared, for example, interfacial polymerization, for example, hydroxymethylcellulose or  
20 gelatin-microcapsules and poly (methylmethacrylate) microcapsules, respectively, or in colloidal drug delivery systems, for example, liposomes, albumin microspheres, microemulsions, nanoparticles and nanocapsules or in macroemulsions. Such techniques are disclosed in Remington's Pharmaceutical Sciences (1980).

EGB 761® and isolated GKB can be administered by oral, parenteral (e.g.,  
25 intramuscular, intraperitoneal, intravenous or subcutaneous injection, or implant), nasal, vaginal, rectal, sublingual or topical routes of administration and can be formulated with pharmaceutically acceptable carriers to provide dosage forms appropriate for each route of administration.

Solid dosage forms for oral administration include capsules, tablets, pills, powders  
30 and granules. In such solid dosage forms, the active compound is admixed with at least one inert pharmaceutically acceptable carrier such as sucrose, lactose, or starch. Such dosage forms can also comprise, as is normal practice, additional substances other than such inert diluents, e.g., lubricating agents such as magnesium stearate. In the case of capsules,

tablets and pills, the dosage forms may also comprise buffering agents. Tablets and pills can additionally be prepared with enteric coatings.

Liquid dosage forms for oral administration include pharmaceutically acceptable emulsions, solutions, suspensions, syrups, the elixirs containing inert diluents commonly  
5 used in the art, such as water. Besides such inert diluents, compositions can also include adjuvants, such as wetting agents, emulsifying and suspending agents, and sweetening, flavoring and perfuming agents.

Preparations according to this invention for parenteral administration include sterile aqueous or non-aqueous solutions, suspensions, or emulsions. Examples of non-aqueous  
10 solvents or vehicles are propylene glycol, polyethylene glycol, vegetable oils, such as olive oil and corn oil, gelatin, and injectable organic esters such as ethyl oleate. Such dosage forms may also contain adjuvants such as preserving, wetting, emulsifying, and dispersing agents. They may be sterilized by, for example, filtration through a bacteria-retaining filter, by incorporating sterilizing agents into the compositions, by irradiating the compositions, or  
15 by heating the compositions. They can also be manufactured in the form of sterile solid compositions which can be dissolved in sterile water, or some other sterile injectable medium immediately before use.

Compositions for rectal or vaginal administration are preferably suppositories which may contain, in addition to the active substance, excipients such as coca butter or a  
20 suppository wax.

Compositions for nasal or sublingual administration are also prepared with standard excipients well known in the art.

The dosage of EGB 761® or isolated GKB in the compositions of this invention may be varied; however, it is necessary that the amount of the active ingredient be such that a  
25 suitable dosage form is obtained. The selected dosage depends upon the desired therapeutic effect, on the route of administration, and on the duration of the treatment. The dose can be administered as a single dose or divided into multiple doses. An effective dose amount of either EGB 761® or isolated GKB depends upon the condition being treated, the route of administration chosen and ultimately will be decided by the attending physician or  
30 veterinarian.

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B1, Ki-67, proliferating cell nuclear antigen, and p34(cdc2). Cancer, 85: 1569-1576, 1999.

The contents of the publications and patents referenced herein are incorporated herein by reference in their entirety.

5

### Claims

What is claimed is:

1. A method of combating cancer in a patient in need of such combating,  
5 wherein the cancer is caused by the deregulation of expression of proteins having a role in regulating tumor cells, which comprises administering an effective amount of Ginkgo biloba extracts or isolated Ginkgolide B to said patient.
2. A method of combating the proliferation of cancer cells in a patient in need of such combating, wherein the proliferation is caused by the deregulation of expression of  
10 proteins having a role in regulating tumor cells, which comprises administering an effective amount of Ginkgo biloba extracts or isolated Ginkgolide B to said patient.
3. A method of combating the proliferation of cancer cells in a patient in need of such combating, wherein the proliferation is caused by over-expression of proteins having a role in regulating tumor cells, which comprises administering an effective amount of Ginkgo  
15 biloba extracts or isolated Ginkgolide B to said patient.
4. A method of combating the proliferation of cancer cells having an aggressive phenotype in a patient in need of such combating, wherein the proliferation is caused by the over-expression of peripheral-type benzodiazepine receptor protein, which comprises administering an effective amount of Ginkgo biloba extracts or isolated Ginkgolide B to said  
20 patient.
5. A method of combating the proliferation of cancer cells, where the proliferation is caused by the over-expression of oncogenes, by decreasing the expression of said oncogenes in a patient in need of such combating, which comprises administering an effective amount of Ginkgo biloba extracts or isolated Ginkgolide B to said patient.
- 25 6. A method according to claim 5, wherein said oncogenes are one or more of APC, PE-1, RhoA and c-Jun.
7. A method of decreasing the expression of peripheral-type benzodiazepine receptor in cancer cells in a patient in need of such decreasing, wherein said cancer cells express an abnormal level of peripheral-type benzodiazepine receptor relative to normal  
30 cancer cells, which comprises administering an effective amount of Ginkgo biloba extracts or isolated Ginkgolide B to said patient.
8. A method according to claim 7, wherein said cancer cells are human breast cancer cells.

9. A method according to claim 7, wherein said cancer cells are glioblastomas.
10. A method according to claim 7, wherein said cancer cells are human brain tumor cells.
11. A method according to claim 7, wherein said cancer cells are human  
5 astrocytoma cells.
12. A method according to claim 7, wherein said cancer cells are human colonic carcinoma cells.
13. A method according to claim 7, wherein said cancer cells are human colonic adenocarcinoma cells.
- 10 14. A method according to claim 7, wherein said cancer cells are human ovarian carcinoma cells.
15. A method according to claim 7, wherein said cancer cells are human hepatocellular carcinoma cells.
16. A method of decreasing the expression of peripheral-type benzodiazepine  
15 receptor mRNA in cancer cells in a patient in need of such decreasing, which comprises administering an effective amount of Ginkgo biloba extracts or isolated Ginkgolide B to said patient.
17. A method of increasing the expression of c-Myc protooncogene in a patient in need of such increasing, which comprises administering an effective amount of Ginkgo  
20 biloba extracts or isolated Ginkgolide B to said patient.
18. A method of decreasing the expression of cell cycle regulators prothymosin- $\alpha$ , CDK2, p53CDC, myeloblastin and p120 proliferating-cell nuclear antigen in a patient in need of such decreasing, which comprises administering an effective amount of Ginkgo biloba extracts or isolated Ginkgolide B to said patient.
- 25 19. A method of decreasing the expression of intracellular signal transduction modulators NET1 and ERK2, in a patient in need of such decreasing, which comprises administering an effective amount of Ginkgo biloba extracts or isolated Ginkgolide B to said patient.
20. A method of decreasing the expression of apoptosis-related products  
30 Adenosine A2A Receptor, Flt3 ligand, Grb2, Clusterin, RXR- $\beta$ , Glutathione S-transferase P, N-Myc, TRADD, SGP-2 and NIP-1, in a patient in need of such decreasing, which comprises

administering an effective amount of Ginkgo biloba extracts or isolated Ginkgolide B to said patient.

21. A method of decreasing the expression of transcription factors Id-2, ATF-4, ETR101 and ETR-103 in a patient in need of such decreasing, which comprises  
5 administering an effective amount of Ginkgo biloba extracts or isolated Ginkgolide B to said patient.

22. A method of decreasing the expression of growth factors macrophage colony-stimulating factor-1, heparin-binding EGF-like growth factor, hepatocyte growth factor-like protein and inhibin  $\alpha$ , in a patient in need of such decreasing, which comprises  
10 administering an effective amount of Ginkgo biloba extracts or isolated Ginkgolide B to said patient.

23. A method of decreasing the expression of cell adhesion molecules CD19 B-lymphocyte antigen, L1CAM,  $\beta$ -catenin, integrin subunits  $\alpha 3$ ,  $\alpha 4$ ,  $\alpha 6$ ,  $\beta 5$ , and  $\alpha M$ , in a patient in need of such decreasing, which comprises administering an effective amount of  
15 Ginkgo biloba extracts or isolated Ginkgolide B to said patient.

24. A method of decreasing the expression of genes APC, PE-1, RhoA, c-Jun, prothymosin- $\alpha$ , CDK2, p53CDC, myeloblastin, p120 proliferating-cell nuclear antigen, NET1, ERK2, Adenosine A2A Receptor, Flt3 ligand, Grb2, Clusterin, RXR- $\beta$ , Glutathione S-transferase P, N-Myc, TRADD, SGP-2, NIP-1, Id-2, ATF-4, ETR-101, ETR-103,  
20 macrophage colony-stimulating factor-1, heparin-binding EGF-like growth factor, hepatocyte growth factor-like protein, inhibin  $\alpha$ , CD19 B-lymphocyte antigen, L1CAM,  $\beta$ -catenin, and integrin subunits  $\alpha 3$ ,  $\alpha 4$ ,  $\alpha 6$ ,  $\beta 5$ , and  $\alpha M$ , in a patient in need of such decreasing, which comprises administering an effective amount of Ginkgo biloba extracts or isolated Ginkgolide B to said patient.

25. 25. A pharmaceutical composition comprising an effective amount of Ginkgo biloba extracts or isolated Ginkgolide B for combating cancer and a pharmaceutically acceptable carrier or diluent.

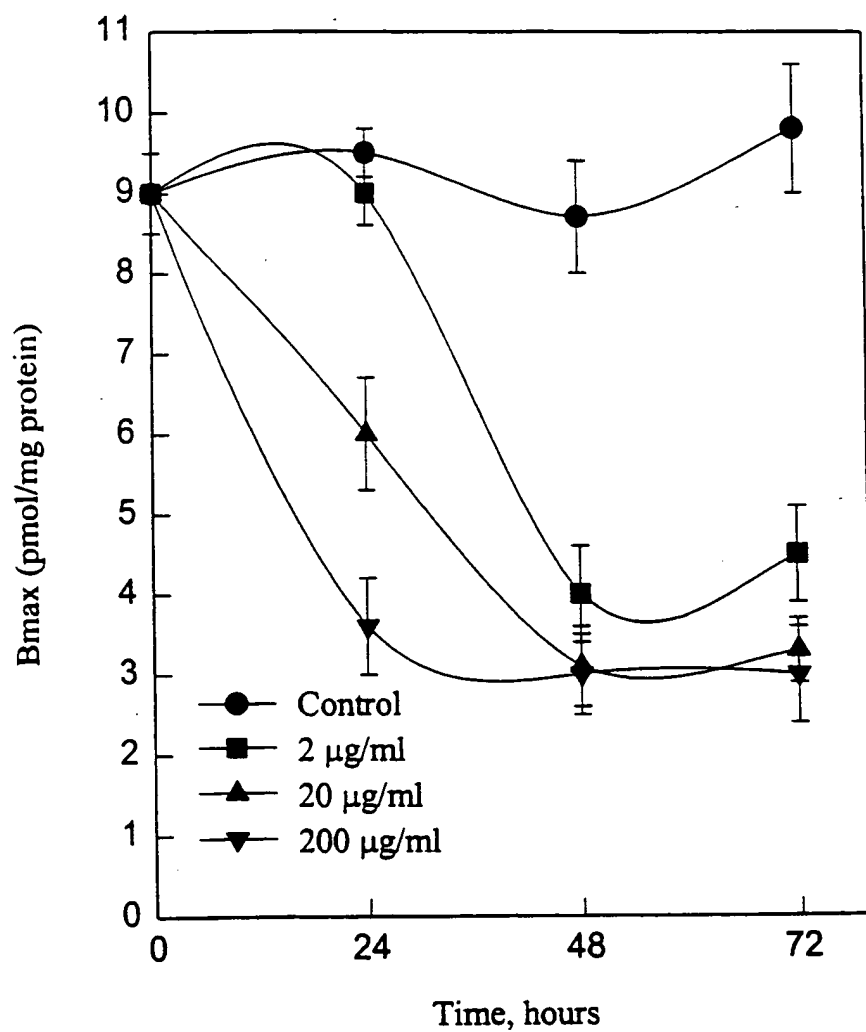


Figure 1

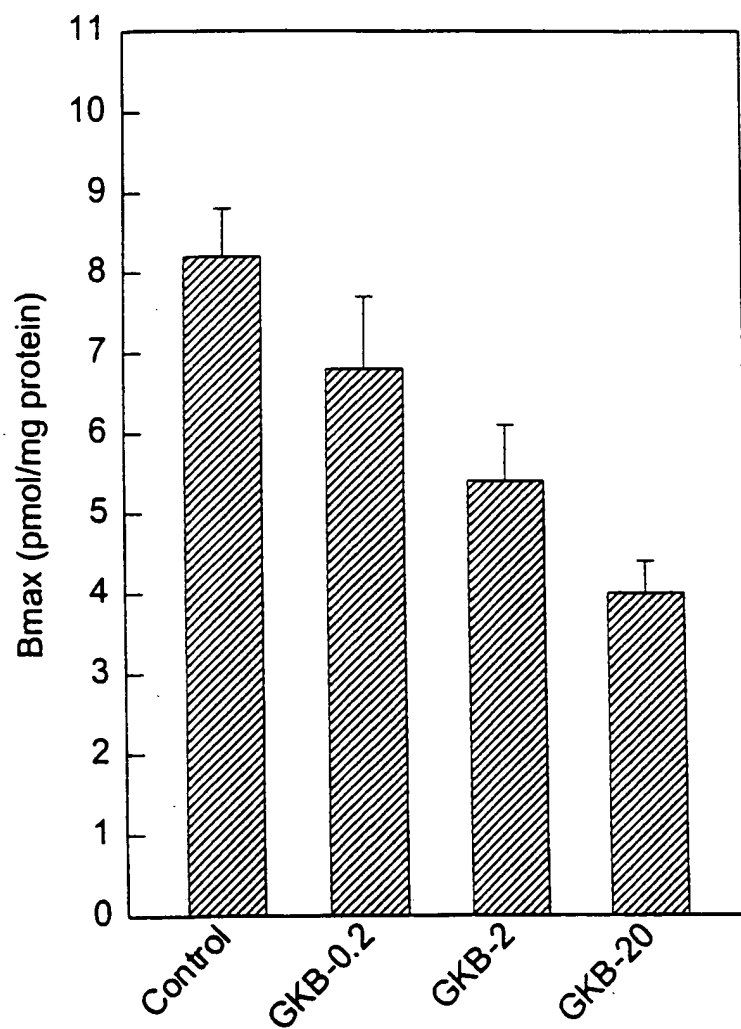


Figure 2

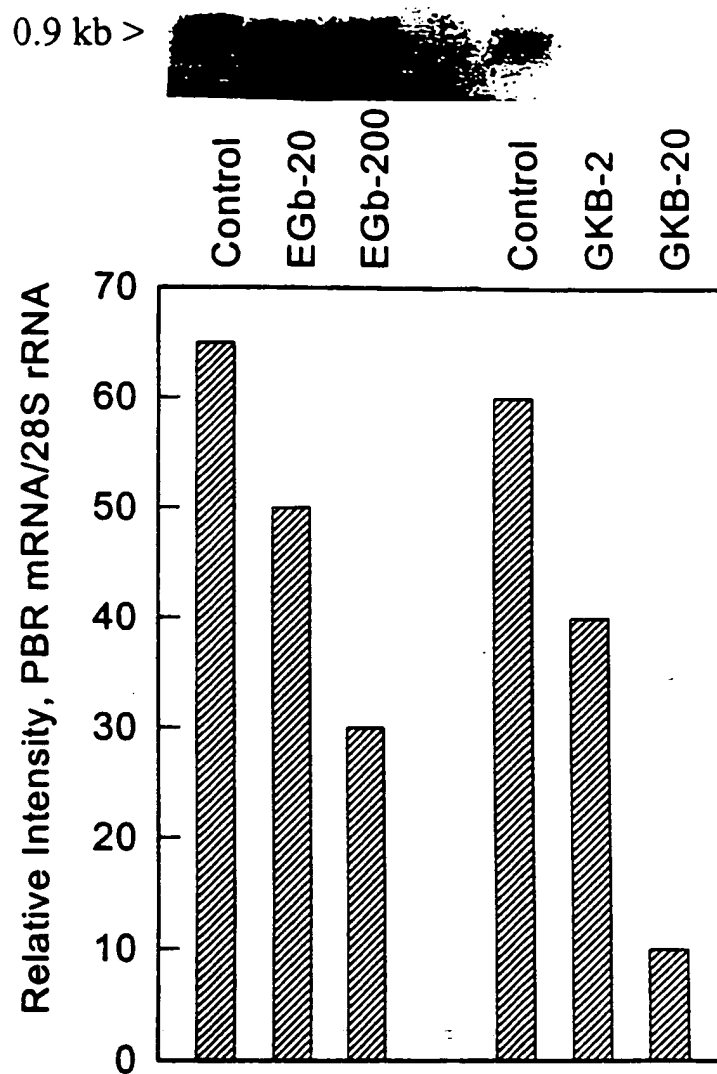


Figure 3

10/018448

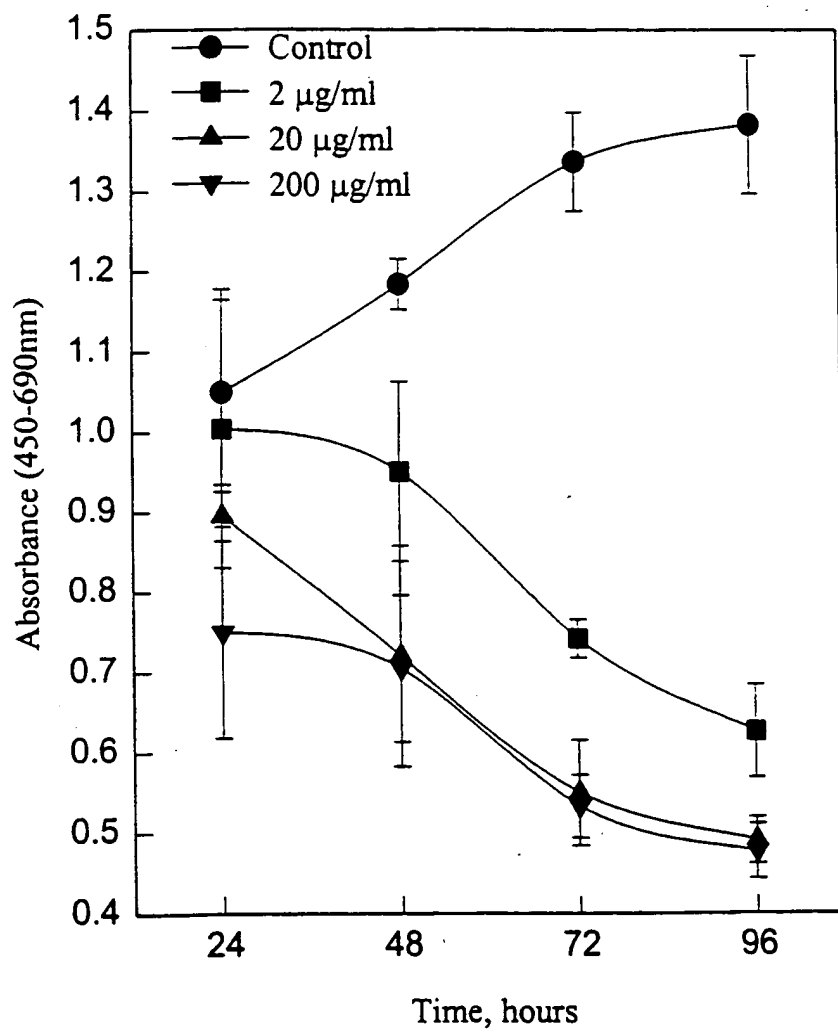


Figure 4



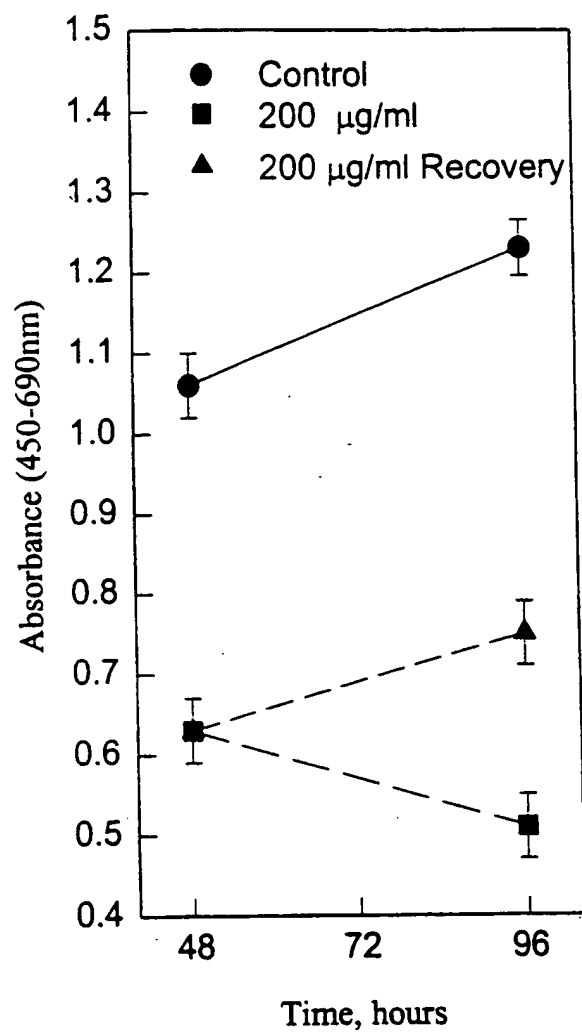


Figure 5: Right

10/018448

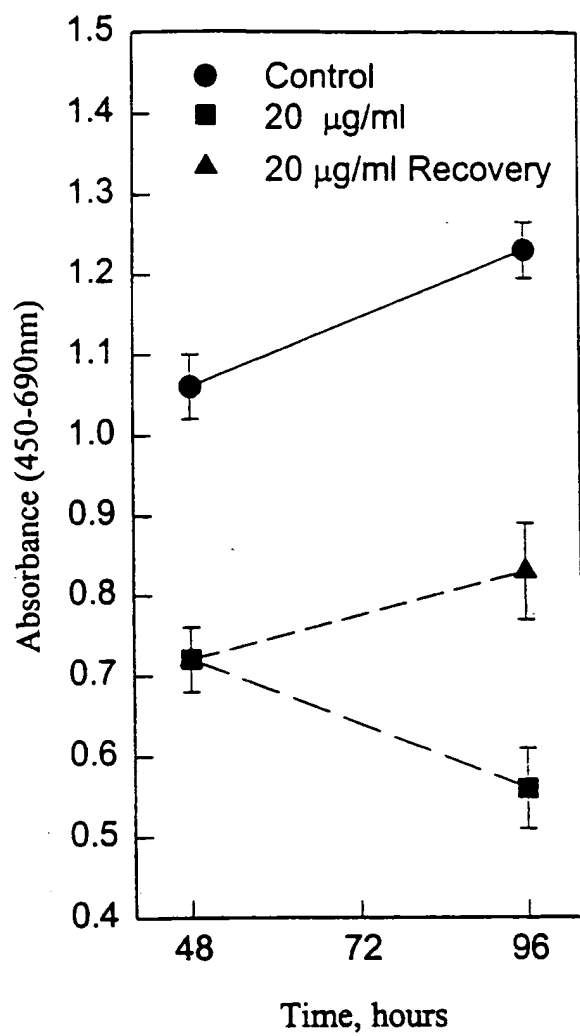


Figure 5: Middle

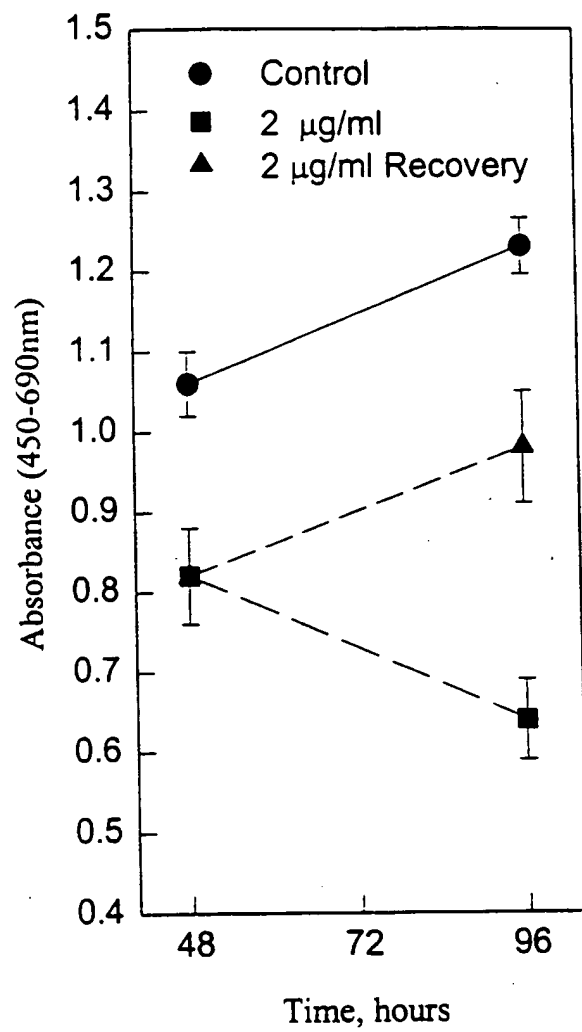


Figure 5: Left

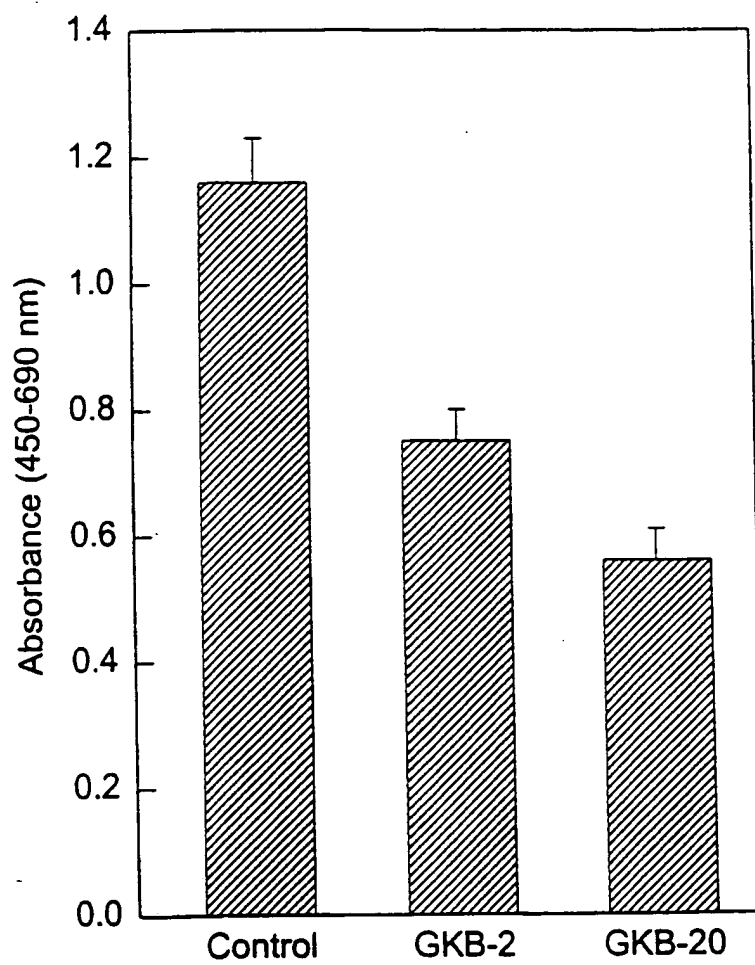


Figure 6

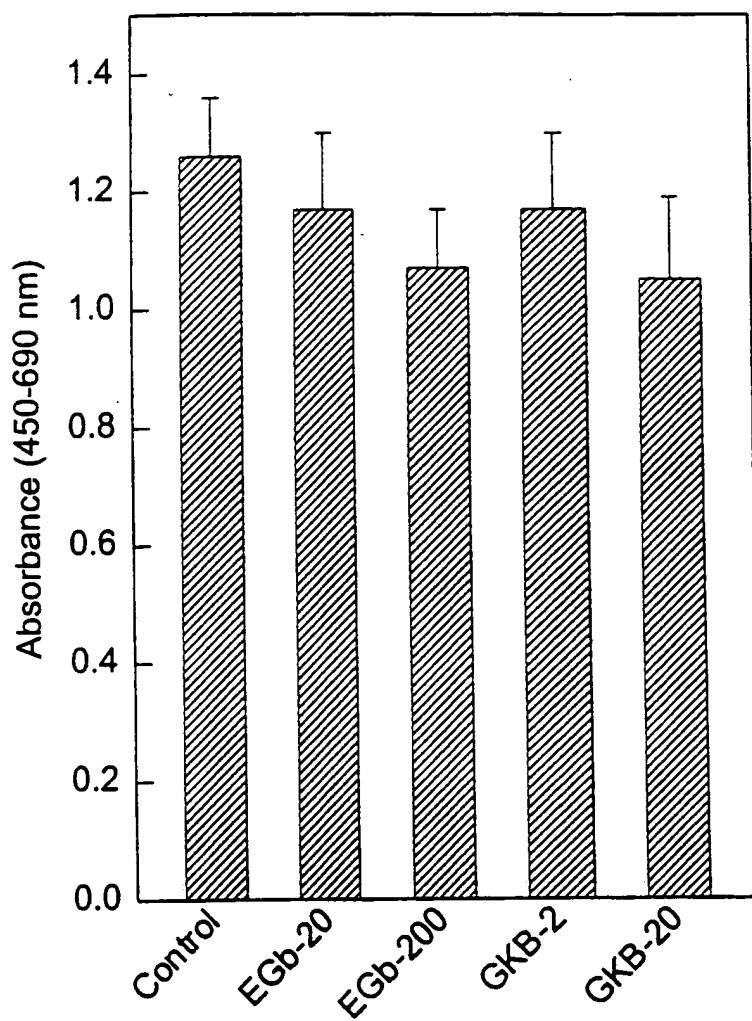


Figure 7

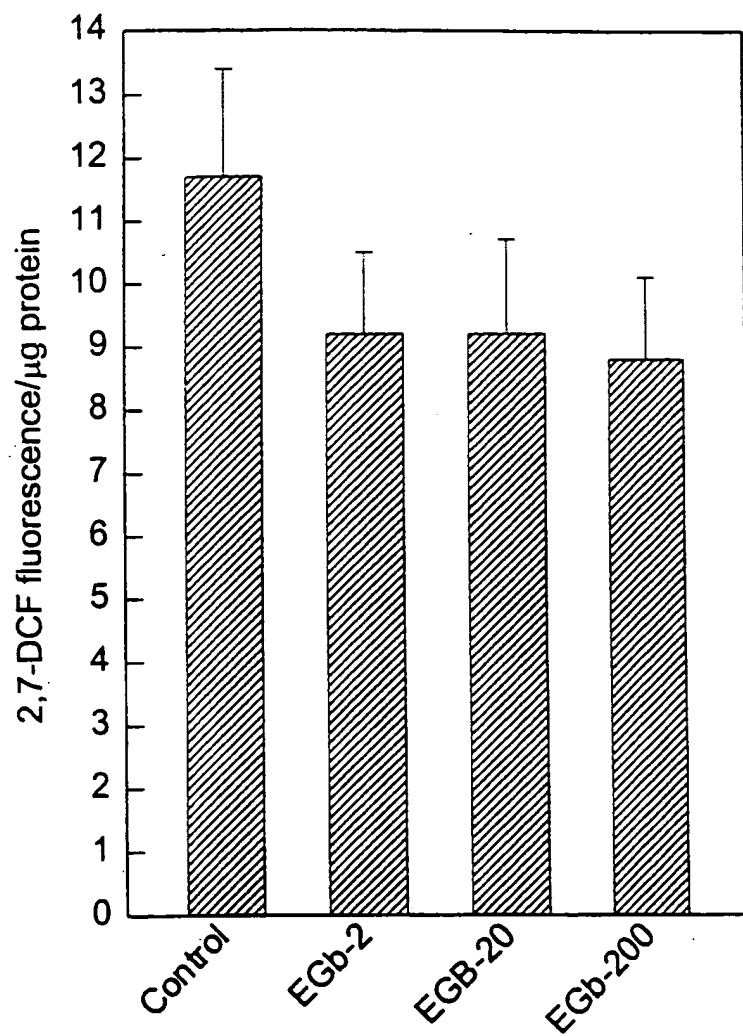


Figure 8

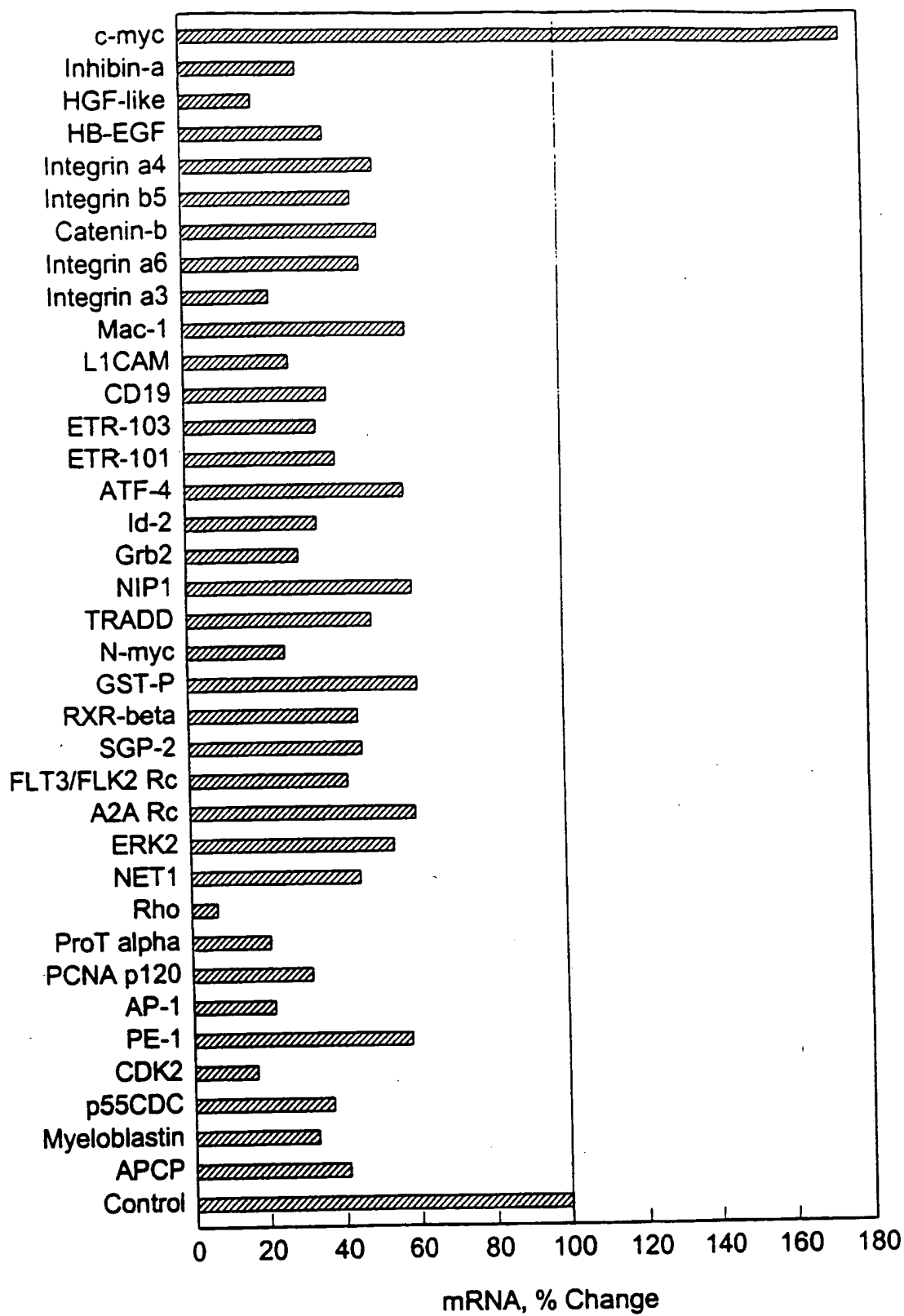


Figure 9

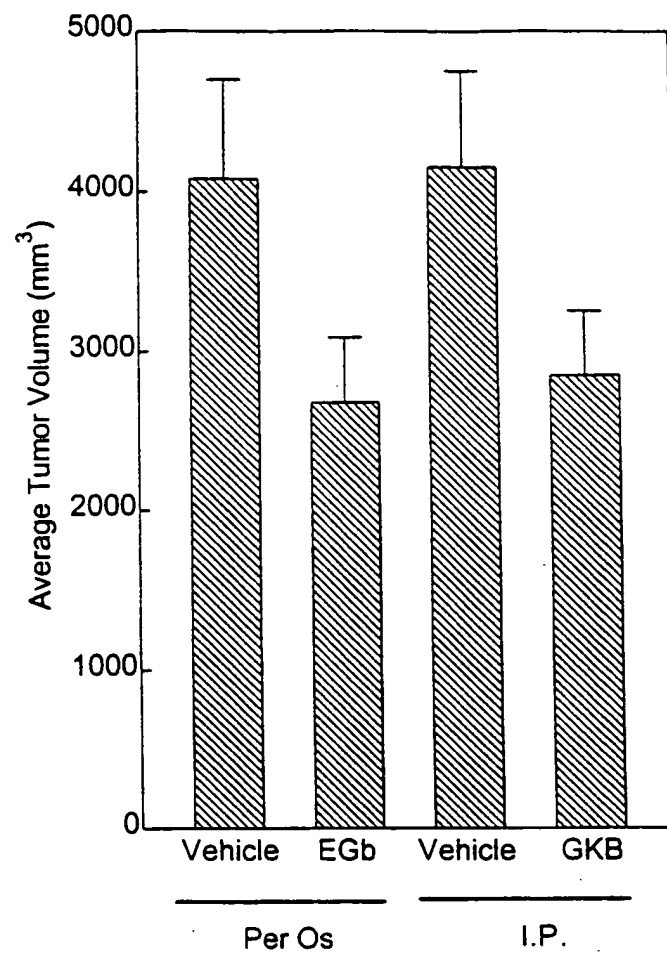


Figure 10



10/018448

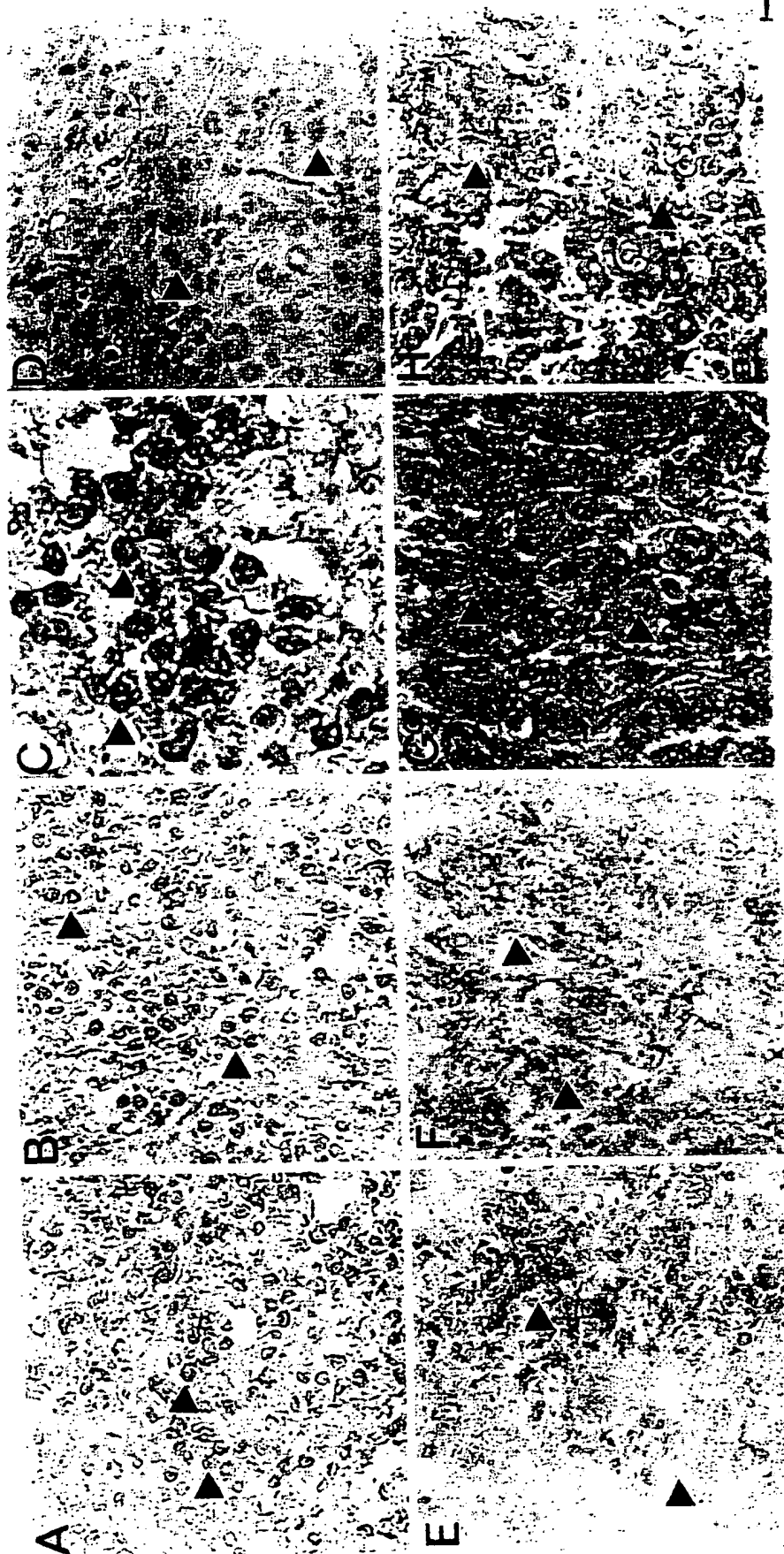


Figure 11

# INTERNATIONAL SEARCH REPORT

Int. Application No  
PCT/US 00/22174

A. CLASSIFICATION OF SUBJECT MATTER  
IPC 7 A61K35/78 A61P35/00

According to International Patent Classification (IPC) or to both national classification and IPC

## B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)  
IPC 7 A61K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

WPI Data, PAJ, EPO-Internal, FSTA, BIOSIS, MEDLINE, PASCAL, LIFESCIENCES, CHEM  
ABS Data, CAB Data, EMBASE

## C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	EP 0 359 951 A (SCHOLLE HELMUT DR MED) 28 March 1990 (1990-03-28) column 1, line 24 - column 2, line 24 ---	1-25
X	DE 42 08 868 A (MICHAELIS PETER DR MED) 9 September 1993 (1993-09-09) column 1, line 3 - line 59 -----	1-25

☐ Further documents are listed in the continuation of box C.

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### \* Special categories of cited documents:

- \*A\* document defining the general state of the art which is not considered to be of particular relevance
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- \*L\* document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)
- \*O\* document referring to an oral disclosure, use, exhibition or other means
- \*P\* document published prior to the international filing date but later than the priority date claimed

\*T\* later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

\*X\* document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

\*Y\* document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.

\*S\* document member of the same patent family

Date of the actual completion of the international search

22 November 2000

Date of mailing of the international search report

29/11/2000

Name and mailing address of the ISA

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Rempp, G

**FURTHER INFORMATION CONTINUED FROM PCT/SA/ 210**

Continuation of Box I.1

Although claims 1-24 are directed to a method of treatment of the human/animal body, the search has been carried out and based on the alleged effects of the compound/composition.

-----

Continuation of Box I.1

Claims Nos.: 1-24

Rule 39.1(iv) PCT - Method for treatment of the human or animal body by therapy

# INTERNATIONAL SEARCH REPORT

information on patent family members

International Application No

PCT/US 00/22174

Patent document cited in search report		Publication date	Patent family member(s)	Publication date
EP 0359951	A	28-03-1990	DE 3832056 A	22-03-1990
			DE 58906809 D	10-03-1994
			ES 2049280 T	16-04-1994
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DE 4208868	A	09-09-1993	NONE	
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(12) INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(19) World Intellectual Property Organization  
International Bureau



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**WO 01/12208 A1**

(51) International Patent Classification<sup>7</sup>: **A61K 35/78,**  
**A61P 35/00**

[FR/FR]: 2, rue de Vouillee, F-75015 Paris (FR). **PA-PADOPOULOS, Vassilios** [US/US]; 15417 Peach Leaf Drive, North Potomac, MD 20878 (US).

(21) International Application Number: PCT/US00/22174

(22) International Filing Date: 11 August 2000 (11.08.2000)

(74) Agent: **TSAO, Y., Rocky**: Fish & Richardson P.C., 225 Franklin Street, Boston, MA 02110-2804 (US).

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60/148,604 12 August 1999 (12.08.1999) US

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(63) Related by continuation (CON) or continuation-in-part (CIP) to earlier application:  
US 60/148,604 (CIP)  
Filed on 12 August 1999 (12.08.1999)

(84) Designated States (*regional*): ARIPO patent (GH, GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG).

(71) Applicants (*for all designated States except US*): **SOCIETE DE CONSEILS DE RECHERCHES ET D'APPLICATIONS SCIENTIFIQUES, S.A.S.** [FR/FR]; 51, 53, rue du Docteur Blanche, F-75016 Paris (FR). **GEORGETOWN UNIVERSITY** [US/US]; 37th and O Streets NW, Washington, DC 20007 (US).

Published:  
— *With international search report.*

(72) Inventors; and

(75) Inventors/Applicants (*for US only*): **DRIEU, Katy**

*For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.*

(54) Title: **USE OF GINKGO EXTRACT**

(57) Abstract: The present invention is directed to the use of the extract of Ginkgo biloba leaves or isolated Ginkgolide B (GKB), a component of the extract of Ginkgo biloba leaves in a method for decreasing the expression of peripheral-type benzodiazepine receptor (PBR) in cells of a patient in need thereof. Further, the present invention is directed to the use of the extract of Ginkgo biloba leaves or isolated GKB in a method for decreasing the proliferation of cancer cells in a patient. More particularly, the present invention is directed to the use of the extract of Ginkgo biloba leaves or isolated GKB in a method of decreasing cancer cell proliferation in a patient wherein the cancer cell is human breast cancer cell. Even more particularly, the present invention is directed to the use of the extract of Ginkgo biloba leaves or isolated GKB in method of decreasing cancer cell proliferation in a patient wherein the cancer cell is of the aggressive and invasive phenotype and expresses high levels of PBR in comparison to non-aggressive cancer cell.

WO 01/12208 A1

# INTERNATIONAL SEARCH REPORT

Inter. Application No

PCT/US 00/22174

**A. CLASSIFICATION OF SUBJECT MATTER**  
 IPC 7 A61K35/78 A61P35/00

According to International Patent Classification (IPC) or to both national classification and IPC

**B. FIELDS SEARCHED**

Minimum documentation searched (classification system followed by classification symbols)

IPC 7 A61K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

WPI Data, PAJ, EPO-Internal, FSTA, BIOSIS, MEDLINE, PASCAL, LIFESCIENCES, CHEM  
 ABS Data, CAB Data, EMBASE

**C. DOCUMENTS CONSIDERED TO BE RELEVANT**

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X	DE 42 08 868 A (MICHAELIS PETER DR MED) 9 September 1993 (1993-09-09) column 1, line 3 - line 59	1-25

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\*X\* document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

\*Y\* document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.

\*G\* document member of the same patent family

Date of the actual completion of the international search

22 November 2000

Date of mailing of the international search report

29/11/2000

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FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

Continuation of Box I.1

Although claims 1-24 are directed to a method of treatment of the human/animal body, the search has been carried out and based on the alleged effects of the compound/composition.

-----

Continuation of Box I.1

Claims Nos.: 1-24

Rule 39.1(iv) PCT - Method for treatment of the human or animal body by therapy

# INTERNATIONAL SEARCH REPORT

Information on patent family members

Inter. Patent Application No

PCT/US 00/22174

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
EP 0359951 A	28-03-1990	DE 3832056 A	22-03-1990
		DE 58906809 D	10-03-1994
		ES 2049280 T	16-04-1994
DE 4208868 A	09-09-1993	NONE	



# PATENT COOPERATION TREATY

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NOTIFICATION OF TRANSMITTAL OF  
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EXAMINATION REPORT  
(PCT Rule 71.1)

Date of mailing  
(day/month/year) 24.10.2001

Applicant's or agent's file reference  
**00537-194WO1**

**IMPORTANT NOTIFICATION**

International application No.  
PCT/US00/22174

International filing date (day/month/year)  
11/08/2000

Priority date (day/month/year)  
12/08/1999

Applicant  
SOCIETE DE CONSEILS DE RECHERCHES ET ... et al.

1. The applicant is hereby notified that this International Preliminary Examining Authority transmits herewith the international preliminary examination report and its annexes, if any, established on the international application.
2. A copy of the report and its annexes, if any, is being transmitted to the International Bureau for communication to all the elected Offices.
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By UXA  
Billing Secretary

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# PATENT COOPERATION TREATY

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(PCT Article 36 and Rule 70)

Applicant's or agent's file reference <b>00537-194WO1</b>	<div style="display: flex; justify-content: space-between;"> <div> <b>FOR FURTHER ACTION</b> </div> <div>           See Notification of Transmittal of International Preliminary Examination Report (Form PCT/IPEA/416)         </div> </div>	
International application No. <b>PCT/US00/22174</b>	International filing date (day/month/year) <b>11/08/2000</b>	Priority date (day/month/year) <b>12/08/1999</b>
International Patent Classification (IPC) or national classification and IPC <b>A61K35/78</b>		
Applicant <b>SOCIETE DE CONSEILS DE RECHERCHES ET ... et al.</b>		
<p>1. This international preliminary examination report has been prepared by this International Preliminary Examining Authority and is transmitted to the applicant according to Article 36.</p> <p>2. This REPORT consists of a total of 7 sheets, including this cover sheet.</p> <p><input type="checkbox"/> This report is also accompanied by ANNEXES, i.e. sheets of the description, claims and/or drawings which have been amended and are the basis for this report and/or sheets containing rectifications made before this Authority (see Rule 70.16 and Section 607 of the Administrative Instructions under the PCT).</p> <p>These annexes consist of a total of    sheets.</p>		
<p>3. This report contains indications relating to the following items:</p> <ul style="list-style-type: none"> <li>I    <input checked="" type="checkbox"/> Basis of the report</li> <li>II   <input type="checkbox"/> Priority</li> <li>III <input checked="" type="checkbox"/> Non-establishment of opinion with regard to novelty, inventive step and industrial applicability</li> <li>IV   <input checked="" type="checkbox"/> Lack of unity of invention</li> <li>V    <input checked="" type="checkbox"/> Reasoned statement under Article 35(2) with regard to novelty, inventive step or industrial applicability; citations and explanations supporting such statement</li> <li>VI   <input type="checkbox"/> Certain documents cited</li> <li>VII <input type="checkbox"/> Certain defects in the international application</li> <li>VIII <input checked="" type="checkbox"/> Certain observations on the international application</li> </ul>		
Date of submission of the demand  <b>12/03/2001</b>	Date of completion of this report  <b>24.10.2001</b>	
Name and mailing address of the international preliminary examining authority:  <div style="display: flex; align-items: center;"> <div>             European Patent Office              D-80298 Munich              Tel. +49 89 2399 - 0 Tx: 523656 epmu d              Fax: +49 89 2399 - 4465           </div> </div>	Authorized officer  <b>Smetankine, L</b>  Telephone No. +49 89 2399 8466	



**INTERNATIONAL PRELIMINARY  
EXAMINATION REPORT**

International application No. PCT/US00/22174

**I. Basis of the report**

1. With regard to the **elements** of the international application (*Replacement sheets which have been furnished to the receiving Office in response to an invitation under Article 14 are referred to in this report as "originally filed" and are not annexed to this report since they do not contain amendments (Rules 70.16 and 70.17)*):

**Description, pages:**

1-30 as originally filed

**Claims, No.:**

1-25 as originally filed

**Drawings, sheets:**

1/13-13/13 as originally filed

2. With regard to the **language**, all the elements marked above were available or furnished to this Authority in the language in which the international application was filed, unless otherwise indicated under this item.

These elements were available or furnished to this Authority in the following language: , which is:

- ☐ the language of a translation furnished for the purposes of the international search (under Rule 23.1(b)).
- ☐ the language of publication of the international application (under Rule 48.3(b)).
- ☐ the language of a translation furnished for the purposes of international preliminary examination (under Rule 55.2 and/or 55.3).

3. With regard to any **nucleotide and/or amino acid sequence** disclosed in the international application, the international preliminary examination was carried out on the basis of the sequence listing:

- ☐ contained in the international application in written form.
- ☐ filed together with the international application in computer readable form.
- ☐ furnished subsequently to this Authority in written form.
- ☐ furnished subsequently to this Authority in computer readable form.
- ☐ The statement that the subsequently furnished written sequence listing does not go beyond the disclosure in the international application as filed has been furnished.
- ☐ The statement that the information recorded in computer readable form is identical to the written sequence listing has been furnished.

4. The amendments have resulted in the cancellation of:

- ☐ the description, pages:
- ☐ the claims, Nos.:

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☐ the drawings, sheets:

5. ☐ This report has been established as if (some of) the amendments had not been made, since they have been considered to go beyond the disclosure as filed (Rule 70.2(c)):

*(Any replacement sheet containing such amendments must be referred to under item 1 and annexed to this report.)*

6. Additional observations, if necessary:

**III. Non-establishment of opinion with regard to novelty, inventive step and industrial applicability**

1. The questions whether the claimed invention appears to be novel, to involve an inventive step (to be non-obvious), or to be industrially applicable have not been examined in respect of:

☐ the entire international application.

☐ claims Nos. .

because:

☒ the said international application, or the said claims Nos. 1-24 relate to the following subject matter which does not require an international preliminary examination (*specify*):  
**see separate sheet**

☐ the description, claims or drawings (*indicate particular elements below*) or said claims Nos. are so unclear that no meaningful opinion could be formed (*specify*):

☐ the claims, or said claims Nos. are so inadequately supported by the description that no meaningful opinion could be formed.

☐ no international search report has been established for the said claims Nos. .

2. A meaningful international preliminary examination cannot be carried out due to the failure of the nucleotide and/or amino acid sequence listing to comply with the standard provided for in Annex C of the Administrative Instructions:

☐ the written form has not been furnished or does not comply with the standard.

☐ the computer readable form has not been furnished or does not comply with the standard.

**IV. Lack of unity of invention**

1. In response to the invitation to restrict or pay additional fees the applicant has:

☐ restricted the claims.

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- ☐ paid additional fees.
- ☐ paid additional fees under protest.
- ☐ neither restricted nor paid additional fees.
2. ☒ This Authority found that the requirement of unity of invention is not complied and chose, according to Rule 68.1, not to invite the applicant to restrict or pay additional fees.
3. This Authority considers that the requirement of unity of invention in accordance with Rules 13.1, 13.2 and 13.3 is
- ☐ complied with.
- ☐ not complied with for the following reasons:
4. Consequently, the following parts of the international application were the subject of international preliminary examination in establishing this report:
- ☒ all parts.
- ☐ the parts relating to claims Nos. .

**V. Reasoned statement under Article 35(2) with regard to novelty, inventive step or industrial applicability; citations and explanations supporting such statement**

1. Statement

Novelty (N)	Yes: Claims
	No: Claims 1-25
Inventive step (IS)	Yes: Claims
	No: Claims 1-25
Industrial applicability (IA)	Yes: Claims 1-25
	No: Claims

2. Citations and explanations  
**see separate sheet**

**VIII. Certain observations on the international application**

The following observations on the clarity of the claims, description, and drawings or on the question whether the claims are fully supported by the description, are made:  
**see separate sheet**

**POINT III:**

Claims 1-24 relate to subject-matter considered by this Authority to be covered by the provisions of Rule 67.1(iv) PCT. Consequently, no opinion will be formulated with respect to the industrial applicability of the subject-matter of these claims (Article 34(4)(a)(i) PCT).

**POINT IV:**

The subject-matter of independent claim 1 to 16 and 25 is already known/not inventive (see the grounds for this objection). The requisite unity of invention (Rule 13.1 PCT) therefore no longer exists inasmuch as a technical relationship involving one or more of the same or corresponding special technical features in the sense of Rule 13.2 PCT does not exist between the subject-matter of the following groups of dependent claims:

1. A pharmaceutical composition comprising Ginkgo biloba extracts or isolated Ginkgolide B for combatting cancer and the treatment of cancer by this composition according to claims 1 to 16 and 25.
2. A method of increasing the expression of c- Myc protooncogene in a patient according to claim 17.
3. A method of decreasing the expression of cell cycle regulators ...in a patient according to claim 18.
4. a method of decreasing the expression of intracellular signal transduction modulators...in a patient according to claim 19.
5. a method of decreasing the expression of apoptosis- related products ... in a patient according to claim 20.
6. a method of decreasing the expression of transcription factors ... in a patient according to claim 21.

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SEPARATE SHEET**

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7. a method of decreasing the expression of growth factors macrophage ....for the treatment of a patient according to claim 22.

8. a method of decreasing the expression of cell adhesion... in a patient according to claim 23.

9. the method of decreasing the expression of genes.... for the treatment of a patient according to claim 24.

According to the Rule 68.1 PCT, the International Examining Authority does not invite the Applicant to restrict the claims or to pay additional fees.

**POINT V:**

EP - A - 0 359 951 (1) - see column 1 line 24 to column 2 line 24 describes the treatment of cancer by Ginkgo biloba extracts (GBE). the mechanism of activity of the cancer treatment belongs to a matter of discovery, which is excluded from the patentability. For these reasons, the subject - matter of claims 1 to 24 seem to lack novelty and inventive step.

A pharmaceutical composition based on GBE and a pharmaceutically acceptable carrier is also described by (1) - see column 1 lines 18 to 26, thus claim 25 seem to lack novelty and inventive step.

DE - A - 42 08 868 (2) - see column 1 lines 3 to 59, describes the use of GBE and those of isolated Ginkgolide B for the treatment of cancer. Thus for the reasons explained in relation with document (1), the subject - matter of claims 1 to 24 seem to lack novelty and inventive step.

A pharmaceutical composition based on GBE and Ginkgolide B are also described - see column 1 lines 12 to 16, thus claim 25 seem to lack novelty.

**POINT VIII:**

The subject - matter of claims 17 to 24 is not clear because the therapeutical treatment

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of the concerned disease is not specified.

Further in claim 24 which is a method of decreasing the expression of genes, in fact concerns others expressions such as growth factor, integrin subunits adenosine receptors...etc, therefore claim 24 is not clear in this respect.



# PATENT COOPERATION TREATY

# PCT

## INTERNATIONAL PRELIMINARY EXAMINATION REPORT

(PCT Article 36 and Rule 70)

REC'D 26 OCT 2001

WPO PCT

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Applicant's or agent's file reference 00537-194WO1		<b>FOR FURTHER ACTION</b> See Notification of Transmittal of International Preliminary Examination Report (Form PCT/IPEA/416)	
International application No. PCT/US00/22174	International filing date (day/month/year) 11/08/2000	Priority date (day/month/year) 12/08/1999	
International Patent Classification (IPC) or national classification and IPC A61K35/78			
Applicant SOCIETE DE CONSEILS DE RECHERCHES ET ... et al.			

1. This international preliminary examination report has been prepared by this International Preliminary Examining Authority and is transmitted to the applicant according to Article 36.



2. This REPORT consists of a total of 7 sheets, including this cover sheet.

- ☐ This report is also accompanied by ANNEXES, i.e. sheets of the description, claims and/or drawings which have been amended and are the basis for this report and/or sheets containing rectifications made before this Authority (see Rule 70.16 and Section 607 of the Administrative Instructions under the PCT).

These annexes consist of a total of sheets.

3. This report contains indications relating to the following items:

- I ☒ Basis of the report
- II ☐ Priority
- III ☒ Non-establishment of opinion with regard to novelty, inventive step and industrial applicability
- IV ☒ Lack of unity of invention
- V ☒ Reasoned statement under Article 35(2) with regard to novelty, inventive step or industrial applicability; citations and explanations supporting such statement
- VI ☐ Certain documents cited
- VII ☐ Certain defects in the international application
- VIII ☒ Certain observations on the international application

Date of submission of the demand 12/03/2001	Date of completion of this report 24.10.2001
Name and mailing address of the international preliminary examining authority:  European Patent Office D-80298 Munich Tel. +49 89 2399 - 0 Tx: 523656 epmu d Fax: +49 89 2399 - 4465	Authorized officer Smetankine, L Telephone No. +49 89 2399 3466 

# INTERNATIONAL PRELIMINARY EXAMINATION REPORT

International application No. PCT/US00/22174

## I. Basis of the report

1. With regard to the **elements** of the international application (*Replacement sheets which have been furnished to the receiving Office in response to an invitation under Article 14 are referred to in this report as "originally filed" and are not annexed to this report since they do not contain amendments (Rules 70.16 and 70.17)*):

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# INTERNATIONAL PRELIMINARY EXAMINATION REPORT

International application No. PCT/US00/22174

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1. The questions whether the claimed invention appears to be novel, to involve an inventive step (to be non-obvious), or to be industrially applicable have not been examined in respect of:

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# INTERNATIONAL PRELIMINARY EXAMINATION REPORT

International application No. PCT/US00/22174

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Novelty (N)	Yes:	Claims	
	No:	Claims	1-25
Inventive step (IS)	Yes:	Claims	
	No:	Claims	1-25
Industrial applicability (IA)	Yes:	Claims	1-25
	No:	Claims	

### 2. Citations and explanations see separate sheet

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see separate sheet

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